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UTILITY PATENT APPLICATION TRANSMITTAL (Only for new nonprovisional applications under 37 C.F.R. § 1.53(b))	Attorney Docket No.	MB1-0010
	First Inventor or Application Identifier	Heard
	Title	Disease-Induced Polynucleotides
	Express Mail Label No.	EK 483897023 US

APPLICATION ELEMENTS See MPEP chapter 600 concerning utility patent application contents.	ADDRESS TO: Assistant Commissioner for Patents Box Patent Application Washington, DC 20231
1. <input checked="" type="checkbox"/> * Fee Transmittal Form (e.g., PTO/SB/17) (Submit an original and a duplicate for fee processing) 2. <input checked="" type="checkbox"/> Specification [Total Pages 29] (preferred arrangement set forth below) - Descriptive title of the Invention - Cross References to Related Applications - Statement Regarding Fed sponsored R & D - Reference to Microfiche Appendix - Background of the Invention - Brief Summary of the Invention - Brief Description of the Drawings (if filed) - Detailed Description - Claim(s) - Abstract of the Disclosure 3. <input checked="" type="checkbox"/> Drawing(s) (35 U.S.C. 113) [Total Sheets 4] 4. Oath or Declaration [Total Pages] a. <input checked="" type="checkbox"/> Newly executed (original or copy) b. <input type="checkbox"/> Copy from a prior application (37 C.F.R. § 1.63(d)) (for continuation/divisional with Box 16 completed) i. <input type="checkbox"/> DELETION OF INVENTOR(S) Signed statement attached deleting inventor(s) named in the prior application, see 37 C.F.R. §§ 1.63(d)(2) and 1.33(b).	5. <input checked="" type="checkbox"/> Microfiche Computer Program (Appendix) 6. Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary) a. <input checked="" type="checkbox"/> Computer Readable Copy b. <input checked="" type="checkbox"/> Paper Copy (identical to computer copy) c. <input checked="" type="checkbox"/> Statement verifying identity of above copies
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DISEASE-INDUCED POLYNUCLEOTIDES

The present invention claims priority in part from US Provisional Application Serial No. 60/125,814 filed March 23, 1999.

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FIELD OF THE INVENTION

This invention is in the field of plant molecular biology and relates to compositions and methods for modifying a plant's traits, in particular plant disease tolerance or resistance.

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BACKGROUND OF THE INVENTION

Gene expression levels are controlled in part at the level of transcription, and transcription is affected by transcription factors. Transcription factors regulate gene expression throughout the life cycle of an organism and so are responsible for differential levels of gene expression at various developmental stages, in different tissue and cell types, and in response to different stimuli. Transcription factors may interact with other proteins or with specific sites on a target gene sequence to activate, suppress or otherwise regulate transcription. In addition, the transcription of the transcription factors themselves may be regulated.

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Because transcription factors are key controlling elements for biological pathways, altering the expression levels of one or more transcription factors may change entire biological pathways in an organism. For example, manipulation of the levels of selected transcription factors may result in increased expression of economically useful proteins or metabolic chemicals in plants or to improve other agriculturally relevant characteristics. Conversely, blocked or reduced expression of a transcription factor may reduce biosynthesis of unwanted compounds or remove an undesirable trait. Therefore, manipulating transcription factor levels in a plant offers tremendous potential in agricultural biotechnology for modifying a plant's traits.

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The present invention provides transcription factors for use in modifying a plant's disease tolerance or resistance.

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SUMMARY OF THE INVENTION

In one aspect, the present invention relates to a transgenic plant comprising a recombinant polynucleotide. The recombinant polynucleotide comprises a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of protein SEQ ID Nos. 2N, where N=1-56. And the presence of the recombinant polynucleotide alters the disease tolerance or resistance

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of the transgenic plant when compared with the same trait of another plant lacking the recombinant polynucleotide.

In one embodiment, the nucleotide sequence encodes a polypeptide comprising a conserved domain which may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain. In a further embodiment, the nucleotide sequence further comprises a promoter operably linked to the nucleotide sequence. The promoter may be a constitutive or inducible or tissue-active.

In a second aspect, the present invention relates to a method for altering a plant's disease tolerance or resistance. The method comprises (a) transforming a plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of protein SEQ ID Nos. 2N, where N=1-56; (b) selecting transformed plants; and (c) identifying a transformed plant with roots having an altered trait.

In one embodiment, the nucleotide sequence encodes a polypeptide comprising a conserved domain which may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain. In a further embodiment, the nucleotide sequence further comprises a promoter operably linked to the nucleotide sequence. The promoter may be a constitutive or inducible or tissue-active.

In a third aspect, the present invention relates to a method for altering the expression levels of at least one gene in a plant. The method comprises (a) transforming the plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of protein SEQ ID Nos. 2N, where N=1-56; and (b) selecting said transformed plant.

In one embodiment, the nucleotide sequence encodes a polypeptide comprising a conserved domain which may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain. In a further embodiment, the nucleotide sequence further comprises a promoter operably linked to the nucleotide sequence. The promoter may be a constitutive or inducible or tissue-active.

In a fourth aspect, the present invention relates to another method for altering the disease tolerance of a plant. The method comprises (a) transforming the plant with a recombinant polynucleotide comprising a nucleotide sequence comprising at least 18 consecutive nucleotides of a sequence selected from the group consisting of SEQ ID Nos. 2N-1, where N= 1-56, and SEQ ID Nos. 113-121; and (b) selecting said transformed plant.

In yet another aspect, the present invention is yet another method for altering a plant's trait. The method comprises (a) providing a database sequence; (b) comparing the database sequence with a polypeptide selected from SEQ ID Nos. 2N, where N= 1-56; (c) selecting a database sequence that meets selected sequence criteria; and (d) transforming

said database sequence in the plant. Alternatively, the database sequence can be compared with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121.

5 In a further aspect, the present invention is a method for altering a plant's trait, and the method entails (a) providing a test polynucleotide; (b) hybridizing the test polynucleotide with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121 at low stringency; and (c) transforming the hybridizing test polynucleotide in a plant to alter a trait of the plant.

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BRIEF DESCRIPTION OF THE FIGURES

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Figures 1a-1e provide a table of exemplary polynucleotide and polypeptide sequences of the invention. The table includes from left to right for each sequence: the SEQ ID No., the internal code reference number, the transcription factor family of the sequence, particular DNA or protein fragments for each sequence, whether the sequence is a polynucleotide or polypeptide sequence, identification of the coding sequence for each full length and identification of any conserved domains for the polypeptide sequences.

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DETAILED DESCRIPTION OF THE INVENTION

DEFINITIONS

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A "recombinant polynucleotide" is a nucleotide sequence comprising a gene coding sequence or a fragment thereof (comprising at least 18 consecutive nucleotides, preferably at least 30 consecutive nucleotides, and more preferably at least 50 consecutive nucleotides). Additionally, the polynucleotide may comprise a promoter, an intron, an enhancer region, a polyadenylation site, a translation initiation site, 5' or 3' untranslated regions, a reporter gene, a selectable marker or the like. The polynucleotide may comprise single stranded or double stranded DNA or RNA. The polynucleotide may comprise modified bases or a modified backbone. The polynucleotide may be genomic, a transcript (such as an mRNA) or a processed nucleotide sequence (such as a cDNA). The polynucleotide may comprise a sequence in either sense or antisense orientations.

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A "recombinant polynucleotide" is a polynucleotide that is not in its native state, e.g., the polynucleotide is comprised of a nucleotide sequence not found in nature or the polynucleotide is separated from nucleotide sequences with which it typically is in proximity or is next to nucleotide sequences with which it typically is not in proximity.

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An "recombinant polypeptide" is a polypeptide derived from the translation of a recombinant polynucleotide or is more enriched in a cell than the polypeptide in its natural

state in a wild type cell, e.g. more than 5% enriched, more than 10% enriched or more than 20% enriched and is not the result of a natural response of a wild type plant or is separated from other components with which it is typically associated with in a cell.

A “transgenic plant” may refer to a plant that contains genetic material not normally found in a wild type plant of the same species, or in a naturally occurring variety or in a cultivar, and which has been introduced into the plant by human manipulation. A transgenic plant is a plant that may contain an expression vector or cassette. The expression cassette comprises a gene coding sequence and allows for the expression of the gene coding sequence. The expression cassette may be introduced into a plant by transformation or by breeding after transformation of a parent plant.

A transgenic plant refers to a whole plant as well as to a plant part, such as seed, fruit, leaf, or root, plant tissue, plant cells or any other plant material, and progeny thereof.

The phrase “altered expression” in reference to polynucleotide or polypeptide expression refers to an expression pattern in the transgenic plant that is different from the expression pattern in the wild type plant or a reference; for example, by expression in a cell type other than a cell type in which the sequence is expressed in the wild type plant, or by expression at a time other than at the time the sequence is expressed in the wild type plant, or by a response to different inducible agents, such as hormones or environmental signals, or at different expression levels (either higher or lower) compared with those found in a wild type plant. The term also refers to lowering the levels of expression to below the detection level or completely abolishing expression. The resulting expression pattern may be transient or stable, constitutive or inducible.

A “transcription factor” (TF) refers to a polynucleotide or polypeptide that controls the expression of a gene or genes either directly by binding to one or more nucleotide sequences associated with a gene coding sequence or indirectly by affecting the level or activity of other polypeptides that do bind directly or indirectly to one or more nucleotide sequences associated with a gene coding sequence. A TF, in this definition, includes any polypeptide that can activate or repress transcription of a single gene or a number of genes. This polypeptide group includes, but is not limited to, DNA binding proteins, protein kinases, protein phosphatases, GTP-binding proteins and receptors.

The transcription factor sequence may comprise a whole coding sequence or a fragment or domain of a coding sequence. A “fragment or domain”, as referred to polypeptides, may be a portion of a polypeptide which performs at least one biological function of the intact polypeptide in substantially the same manner or to a similar extent as does the intact polypeptide. A fragment may comprise, for example, a DNA binding domain that binds to a specific DNA promoter region, an activation domain or a domain for protein-protein interactions. Fragments may vary in size from as few as 6 amino acids to the length of the

intact polypeptide, but are preferably at least 30 amino acids in length and more preferably at least 60 amino acids in length. In reference to a nucleotide sequence “a fragment” refers to any sequence of at least consecutive 18 nucleotides, preferably at least 30 nucleotides, more preferably at least 50, of any of the sequences provided herein. Exemplary polynucleotides or

5 polypeptides comprise a sequence provided in the Sequence Listing as SEQ ID No.1 (G1043), SEQ ID No.2 (G1043 protein), SEQ ID No.3 (G759), SEQ ID No.4 (G759 protein), SEQ ID No.5 (G185), SEQ ID No.6 (G185 protein), SEQ ID No.7 (G629), SEQ ID No.8 (G629 protein), SEQ ID No.9 (G435), SEQ ID No.10 (G435 protein), SEQ ID No.11 (G4), SEQ ID No.12 (G4 protein), SEQ ID No.13 (G1035), SEQ ID No.14 (G1035 protein), SEQ ID No.15 (G179), SEQ ID No.16 (G179 protein), SEQ ID No.17 (G28), SEQ ID No.18 (G28 protein), SEQ ID No.19 (G1241), SEQ ID No.20 (G1241 protein), SEQ ID No.21 (G19), SEQ ID No.22 (G19 protein), SEQ ID No.23 (G503), SEQ ID No.24 (G503 protein), SEQ ID No.25 (G263), SEQ ID No.26 (G263 protein), SEQ ID No.27 (G921), SEQ ID No.28 (G921 protein), SEQ ID No.29 (G1275), SEQ ID No.30 (G1275 protein), SEQ ID No.31 (G242), SEQ ID No.32 (G242 protein), SEQ ID No.33 (G1006), SEQ ID No.34 (G1006 protein), SEQ ID No.35 (G1049), SEQ ID No.36 (G1049 protein), SEQ ID No.37 (G502), SEQ ID No.38 (G502 protein), SEQ ID No.39 (G239), SEQ ID No.40 (G239 protein), SEQ ID No.41 (G555), SEQ ID No.42 (G555 protein), SEQ ID No.43 (G352), SEQ ID No.44 (G352 protein), SEQ ID No.45 (G1352), SEQ ID No.46 (G1352 protein), SEQ ID No.47 (G1089), SEQ ID No.48 (G1089 protein), SEQ ID No.49 (G553), SEQ ID No.50 (G553 protein), SEQ ID No.51 (G1221), SEQ ID No.52 (G1221 protein), SEQ ID No.53 (G580), SEQ ID No.54 (G580 protein), SEQ ID No.55 (G270), SEQ ID No.56 (G270 protein), SEQ ID No.57 (G201), SEQ ID No.58 (G201 protein), SEQ ID No.59 (G1417), SEQ ID No.60 (G1417 protein), SEQ ID No.61 (G233), SEQ ID No.62 (G233 protein), SEQ ID No.63 (G920), SEQ ID No.64 (G920 protein), SEQ ID No.65 (G867), SEQ ID No.66 (G867 protein), SEQ ID No.67 (G659), SEQ ID No.68 (G659 protein), SEQ ID No.69 (G620), SEQ ID No.70 (G620 protein), SEQ ID No.71 (G596), SEQ ID No.72 (G596 protein), SEQ ID No.73 (G511), SEQ ID No.74 (G511 protein), SEQ ID No.75 (G471), SEQ ID No.76 (G471 protein), SEQ ID No.77 (G385), SEQ ID No.78 (G385 protein), SEQ ID No.79 (G261), SEQ ID No.80 (G261 protein), SEQ ID No.81 (G25), SEQ ID No.82 (G25 protein), SEQ ID No.83 (G610), SEQ ID No.84 (G610 protein), SEQ ID No.85 (G229), SEQ ID No.86 (G229 protein), SEQ ID No.87 (G221), SEQ ID No.88 (G221 protein), SEQ ID No.89 (G186), SEQ ID No.90 (G186 protein), SEQ ID No.91 (G562), SEQ ID No.92 (G562 protein), SEQ ID No.93 (G255), SEQ ID No.94 (G255 protein), SEQ ID No.95 (G3), SEQ ID No.96 (G3 protein), SEQ ID No.97 (G713), SEQ ID No.98 (G713 protein), SEQ ID No.99 (G515), SEQ ID No.100 (G515 protein), SEQ ID No.101 (G390), SEQ ID No.102 (G390 protein), SEQ ID No.103 (G1034), SEQ ID No.104 (G1034 protein), SEQ ID No.105 (G1149), SEQ ID No.106 (G1149 protein), SEQ ID No.107 (G1334), SEQ ID No.108 (G1334 protein), SEQ ID No.109 (G1650), SEQ ID

No.110 (G1650 protein), SEQ ID No.111 (G241), SEQ ID No.112 (G241 protein), SEQ ID No.113 (G348), SEQ ID No.114 (G171), SEQ ID No.115 (G521), SEQ ID No.116 (G1274), SEQ ID No.117 (G182), SEQ ID No.118 (G1290), SEQ ID No.119 (G374), SEQ ID No.120 (G682) and SEQ ID No.121 (G501).

5 A “conserved domain” refers to a polynucleotide or polypeptide fragment that is more conserved at a sequence level than other fragments when the polynucleotide or polypeptide is compared with homologous genes or proteins from other plants. The conserved domain may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain.

10 A nucleotide sequence is “operably linked” when it is placed into a functional relationship with another nucleotide sequence. For example, a promoter or enhancer is operably linked to a gene coding sequence if the presence of the promoter or enhancer increases the level of expression of the gene coding sequence.

15 “Trait” refers to a physiological, morphological, biochemical or physical characteristic of a plant or particular plant material or cell. This characteristic may be visible to the human eye, such as seed or plant size, or be measured by biochemical techniques, such as the protein, starch or oil content of seed or leaves or by the observation of the expression level of genes by employing Northern, RT PCR, microarray gene expression assays or reporter gene expression systems or be measured by agricultural observations such as stress tolerance, yield or disease resistance.

20 “Trait modification” refers to a detectable difference in a characteristic in a transgenic plant expressing a polynucleotide or polypeptide of the present invention relative to a plant not doing so, such as a wild type plant. The trait modification may entail at least a 5% increase or decrease in an observed trait (difference), at least a 10% difference, at least a 20% difference, at least a 30%, at least a 50%, at least a 70%, at least a 100% or a greater difference. It is known that there may be a natural variation in the modified trait. Therefore, the trait modification observed entails a change in the normal distribution of the trait in transgenic plants compared with the distribution observed in wild type plant.

25 Trait modifications of particular interest include those to seed (embryo), fruit, root, flower, leaf, stem, shoot, seedling or the like, including: enhanced tolerance to environmental conditions including freezing, chilling, heat, drought, water saturation, radiation and ozone; enhanced resistance to microbial, fungal or viral diseases; resistance to nematodes, decreased herbicide sensitivity, enhanced tolerance of heavy metals (or enhanced ability to take up heavy metals), enhanced growth under poor photoconditions (e.g., low light and/or short day length), or changes in expression levels of genes of interest. Other phenotypes that may be modified relate to the production of plant metabolites, such as variations in the production of taxol, tocopherol, tocotrienol, sterols, phytosterols, vitamins, wax monomers,

anti-oxidants, amino acids, lignins, cellulose, tannins, prenyllipids (such as chlorophylls and carotenoids), glucosinolates, and terpenoids, enhanced or compositionally altered protein or oil production (especially in seeds), or modified sugar (insoluble or soluble) and/or starch composition. Physical plant characteristics that may be modified include cell development (such as the number of trichomes), fruit and seed size and number, yields of plant parts such as stems, leaves and roots, the stability of the seeds during storage, characteristics of the seed pod (e.g., susceptibility to shattering), root hair length and quantity, internode distances, or the quality of seed coat. Plant growth characteristics that may be modified include growth rate, germination rate of seeds, vigor of plants and seedlings, leaf and flower senescence, male sterility, apomixis, flowering time, flower abscission, rate of nitrogen uptake, biomass or transpiration characteristics, as well as plant architecture characteristics such as apical dominance, branching patterns, number of organs, organ identity, organ shape or size.

Of particular interest are traits relating to increased disease resistance or tolerance of a plant, such as alterations in cell wall composition, trichome number or structure, callose induction, phytoalexin induction, alterations in the cell death response or the like. These transgenic plants may be more resistant to biotrophic or necrotrophic pathogens such as a fungus, bacterium, mollicute, virus, nematode, a parasitic higher plant or the like and associated diseases. Another desirable phenotype is a change in the overall gene expression pattern of the plant in response to disease.

1. The Sequences

We have discovered particular plant transcription factors (TFs) that are induced when plants are exposed to either biotrophic or necrotrophic pathogens.. These transgenic plants may be more resistant to biotrophic or necrotrophic pathogens such as a fungus, bacterium, mollicute, virus, nematode, a parasitic higher plant or the like and associated diseases, in particular, pathogens such as *Fusarium oxysporum*, *Erysiphe orontii* and other powdery mildews, *Sclerotinia spp.*, soil-borne oomycetes, foliar oomycetes, *Botrytis spp.*, *Rhizoctonia spp.*, *Verticillium dahliae/albo-atrum*, *Alternaria spp.*, rusts, *Mycosphaerella spp.*, *Fusarium solani*, or the like. The diseases include fungal diseases such as rusts, smuts, wilts, yellows, root rot, leaf drop, ergot, leaf blight of potato, brown spot of rice, leaf blight, late blight, powdery mildew, downy mildew, and the like; viral diseases such as sugarcane mosaic, cassava mosaic, sugar beet yellows, plum pox, barley yellow dwarf, tomato yellow leaf curl, tomato spotted wilt virus, and the like; bacterial diseases such as citrus canker, bacterial leaf blight, bacterial wilt, soft rot of vegetables, and the like; nematode diseases such as root knot, sugar beet cyst nematode or the like.

These transcription factors can be used to modulate a plant's response to disease. The plant transcription factors may belong to one of the following transcription factor families:

the AP2 (APETALA2) domain transcription factor family (Riechmann and Meyerowitz (1998) *J. Biol. Chem.* 379:633-646); the MYB transcription factor family (Martin and Paz-Ares, (1997) *Trends Genet.* 13:67-73); the MADS domain transcription factor family (Riechmann and Meyerowitz (1997) *J. Biol. Chem.* 378:1079-1101); the WRKY protein family (Ishiguro and Nakamura (1994) *Mol. Gen. Genet.* 244:563-571); the ankyrin-repeat protein family (Zhang et al. (1992) *Plant Cell* 4:1575-1588); the zinc finger protein (Z) family (Klug and Schwabe (1995) *FASEB J.* 9: 597-604); the homeobox (HB) protein family (Duboule (1994) *Guidebook to the Homeobox Genes*, Oxford University Press); the CAAT-element binding proteins (Forsburg and Guarente (1989) *Genes Dev.* 3:1166-1178); the squamosa promoter binding proteins (SPB) (Klein et al. (1996) *Mol. Gen. Genet.* 1996 250:7-16); the NAM protein family (Souer et al. (1996) *Cell* 85:159-170); the IAA/AUX proteins (Rouse et al. (1998) *Science* 279:1371-1373); the HLH/MYC protein family (Littlewood et al. (1994) *Prot. Profile* 1:639-709); the DNA-binding protein (DBP) family (Tucker et al. (1994) *EMBO J.* 13:2994-3002); the bZIP family of transcription factors (Foster et al. (1994) *FASEB J.* 8:192-200); the Box P-binding protein (the BPF-1) family (da Costa e Silva et al. (1993) *Plant J.* 4:125-135); the high mobility group (HMG) family (Bustin and Reeves (1996) *Prog. Nucl. Acids Res. Mol. Biol.* 54:35-100); the scarecrow (SCR) family (Di Laurenzio et al. (1996) *Cell* 86:423-433); the GF14 family (Wu et al. (1997) *Plant Physiol.* 114:1421-1431); the polycomb (PCOMB) family (Kennison (1995) *Annu. Rev. Genet.* 29:289-303); the teosinte branched (TEO) family (Luo et al. (1996) *Nature* 383:794-799); the ABI3 family (Giraudat et al. (1992) *Plant Cell* 4:1251-1261); the triple helix (TH) family (Dehesh et al. (1990) *Science* 250:1397-1399); the EIL family (Chao et al. (1997) *Cell* 89:1133-44); the AT-HOOK family (Reeves and Nissen (1990) *Journal of Biological Chemistry* 265:8573-8582); the S1FA family (Zhou et al. (1995) *Nucleic Acids Res.* 23:1165-1169); the bZIPT2 family (Lu and Ferl (1995) *Plant Physiol.* 109:723); the YABBY family (Bowman et al. (1999) *Development* 126:2387-96); the PAZ family (Bohmert et al. (1998) *EMBO J.* 17:170-80); a family of miscellaneous (MISC) transcription factors including the DPBF family (Kim et al. (1997) *Plant J.* 11:1237-1251) and the SPF1 family (Ishiguro and Nakamura (1994) *Mol. Gen. Genet.* 244:563-571); the golden (GLD) family (Hall et al. (1998) *Plant Cell* 10:925-936).

Producing transgenic plants with modified expression levels of one or more of these transcription factors compared with those levels found in a wild type or reference plant may be used to modify a plant's traits. The effect of modifying the expression levels of a particular transcription factor on the traits of a transgenic plant is described further in the Examples.

The polynucleotides and polypeptides are provided in the Sequence Listing and are tabulated in Figure 1. Figure 1 identifies a SEQ ID No., its corresponding GID number, the transcription factor family to which the sequence belongs, fragments derived from the sequences, whether the sequence is a polynucleotide or a polypeptide sequence, the full

length coding sequences and conserved domains. We have also identified domains or fragments derived from the sequences. The numbers indicating the fragment location for the DNA sequences may be from either 5' or 3' end of the DNA. For the protein sequences the fragment location is determined from the N-terminus of the protein and may include adjacent amino acid sequences, such as for example for SEQ ID No. 2 an additional 10, 20, 40, 60 or 100 amino acids in either N-terminal or C-terminal direction of the described fragments.

The identified polypeptide fragments may be linked to fragments or sequences derived from other transcription factors so as to generate additional novel sequences, such as by employing the methods described in Short, PCT publication WO9827230, entitled "Methods and Compositions for Polypeptide Engineering" or in Patten et al., PCT publication WO9923236, entitled "Method of DNA Shuffling". Alternatively, the identified fragment may be linked to a transcription activation domain. A transcription activation domain assists in initiating transcription from a DNA binding site. A common feature of some activation domains is that they are designed to form amphiphilic alpha helices with excess positive or negative charge (Giniger and Ptashne (1987) Nature 330:670-672, Gill and Ptashne (1987) Cell 51:121-126, Estruch et al (1994) Nucl. Acids Res. 22:3983-3989). Examples include the transcription activation region of VP16 or GAL4 (Moore et al. (1998) Proc. Natl. Acad. Sci. USA 95: 376-381; and Aoyama et al. (1995) Plant Cell 7:1773-1785), peptides derived from bacterial sequences (Ma and Ptashne (1987) Cell 51: 113-119) and synthetic peptides (Giniger and Ptashne, supra).

The isolated polynucleotides and polypeptides may be used to modify plant development, physiology or biochemistry such that the modified plants have a trait advantage over wild type plants. The identified polynucleotide fragments are also useful as nucleic acid probes and primers. A nucleic acid probe is useful in hybridization protocols, including protocols for microarray experiments. Primers may be annealed to a complementary target DNA strand by nucleic acid hybridization to form a hybrid between the primer and the target DNA strand, and then extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR) or other nucleic-acid amplification methods. See Sambrook et al., *Molecular Cloning. A Laboratory Manual*, Ed. 2, Cold Spring Harbor Laboratory Press, New York (1989) and Ausubel et al. (eds) *Current Protocols in Molecular Biology*, John Wiley & Sons (1998).

2. Identification of Homologous Sequences (Homologs)

Homologous sequences to those provided in the Sequence Listing derived from *Arabidopsis thaliana* or from other plants may be used to modify a plant trait. Homologous sequences may be derived from any plant including monocots and dicots and in particular

agriculturally important plant species, including but not limited to, crops such as soybean, wheat, corn, potato, cotton, rice, oilseed rape (including canola), sunflower, alfalfa, sugarcane and turf; or fruits and vegetables, such as banana, blackberry, blueberry, strawberry, and raspberry, cantaloupe, carrot, cauliflower, coffee, cucumber, eggplant, grapes, honeydew, lettuce, mango, melon, onion, papaya, peas, peppers, pineapple, spinach, squash, sweet corn, tobacco, tomato, watermelon, rosaceous fruits (such as apple, peach, pear, cherry and plum) and vegetable brassicas (such as broccoli, cabbage, cauliflower, brussel sprouts and kohlrabi). Other crops, fruits and vegetables whose phenotype may be changed include barley, currant, avocado, citrus fruits such as oranges, lemons, grapefruit and tangerines, artichoke, cherries, nuts such as the walnut and peanut, endive, leek, roots, such as arrowroot, beet, cassava, turnip, radish, yam, sweet potato and beans. The homologs may also be derived from woody species, such as pine, poplar and eucalyptus.

Substitutions, deletions and insertions introduced into the sequences provided in the Sequence Listing are also envisioned by the invention. Such sequence modifications can be engineered into a sequence by site-directed mutagenesis (Wu (ed.) *Meth. Enzymol.* (1993) vol. 217, Academic Press). Amino acid substitutions are typically of single residues; insertions usually will be on the order of about from 1 to 10 amino acid residues; and deletions will range about from 1 to 30 residues. In preferred embodiments, deletions or insertions are made in adjacent pairs, e.g., a deletion of two residues or insertion of two residues. Substitutions, deletions, insertions or any combination thereof may be combined to arrive at a sequence. The mutations that are made in the polynucleotide encoding the transcription factor should not place the sequence out of reading frame and should not create complementary regions that could produce secondary mRNA structure.

Substitutions are those in which at least one residue in the amino acid sequence has been removed and a different residue inserted in its place. Such substitutions may be conservative with little effect on the function of the gene, for example by substituting alanines for serines, arginines for lysines, glutamate for aspartate and the like. The substitutions which are not conservative are expected to produce the greatest changes in protein properties will be those in which (a) a hydrophilic residue, e.g., seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g., leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine.

Additionally, the term "homologous sequence" may encompass a polypeptide sequence that is modified by chemical or enzymatic means. The homologous sequence may be a sequence modified by lipids, sugars, peptides, organic or inorganic compounds, by the

use of modified amino acids or the like. Protein modification techniques are illustrated in Ausubel et al. (eds) *Current Protocols in Molecular Biology*, John Wiley & Sons (1998).

Homologous sequences also may mean two sequences having a substantial percentage of sequence identity after alignment as determined by using sequence analysis programs for database searching and sequence alignment and comparison available, for example, from the Wisconsin Package Version 10.0, such as BLAST, FASTA, PILEUP, FINDPATTERNS or the like (GCG, Madison, WI). Public sequence databases such as GenBank, EMBL, Swiss-Prot and PIR or private sequence databases such as PhytoSeq (Incyte Pharmaceuticals, Palo Alto, CA) may be searched. Alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman (1981) *Adv. Appl. Math.* 2:482, by the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. U.S.A.* 85: 2444, by computerized implementations of these algorithms. After alignment, sequence comparisons between two (or more) polynucleotides or polypeptides are typically performed by comparing sequences of the two sequences over a comparison window to identify and compare local regions of sequence similarity. The comparison window may be a segment of at least about 20 contiguous positions, usually about 50 to about 200, more usually about 100 to about 150 contiguous positions. A description of the method is provided in Ausubel et al. (eds) (1999) *Current Protocols in Molecular Biology*, John Wiley & Sons.

Transcription factors that are homologs of the disclosed sequences will typically share at least 40% amino acid sequence identity. More closely related TFs may share at least 50%, 60%, 65%, 70%, 75% or 80% sequence identity with the disclosed sequences. Factors that are most closely related to the disclosed sequences share at least 85%, 90% or 95% sequence identity. At the nucleotide level, the sequences will typically share at least 40% nucleotide sequence identity, preferably at least 50%, 60%, 70% or 80% sequence identity, and more preferably 85%, 90%, 95% or 97% sequence identity. The degeneracy of the genetic code enables major variations in the nucleotide sequence of a polynucleotide while maintaining the amino acid sequence of the encoded protein.

One way to identify whether two nucleic acid molecules are closely related is that the two molecules hybridize to each other under stringent conditions. Generally, stringent conditions are selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Conditions for nucleic acid hybridization and calculation of stringencies can be found in Sambrook et al. (1989) *Molecular Cloning. A Laboratory Manual*, Ed. 2, Cold Spring Harbor Laboratory Press, New York and Tijssen (1993) *Laboratory Techniques in Biochemistry and*

Molecular Biology--Hybridization with Nucleic Acid Probes Part I, Elsevier, New York . Nucleic acid molecules that hybridize under stringent conditions will typically hybridize to a probe based on either the entire cDNA or selected portions of the cDNA under wash conditions of 0.2x SSC to 2.0 x SSC, 0.1% SDS at 50-65° C, for example 0.2 x SSC, 0.1% SDS at 65° C. For detecting less closely related homologs washes may be performed at 50° C.

For conventional hybridization the hybridization probe is conjugated with a detectable label such as a radioactive label, and the probe is preferably of at least 20 nucleotides in length. As is well known in the art, increasing the length of hybridization probes tends to give enhanced specificity. The labeled probe derived from the *Arabidopsis* nucleotide sequence may be hybridized to a plant cDNA or genomic library and the hybridization signal detected using means known in the art. The hybridizing colony or plaque (depending on the type of library used) is then purified and the cloned sequence contained in that colony or plaque isolated and characterized. Homologs may also be identified by PCR-based techniques, such as inverse PCR or RACE, using degenerate primers. See Ausubel et al. (eds) (1998) *Current Protocols in Molecular Biology*, John Wiley & Sons.

TF homologs may alternatively be obtained by immunoscreening an expression library. With the provision herein of the disclosed TF nucleic acid sequences, the polypeptide may be expressed and purified in a heterologous expression system (e.g., *E. coli*) and used to raise antibodies (monoclonal or polyclonal) specific for the TF. Antibodies may also be raised against synthetic peptides derived from TF amino acid sequences. Methods of raising antibodies are well known in the art and are described in Harlow and Lane (1988) *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, New York. Such antibodies can then be used to screen an expression library produced from the plant from which it is desired to clone the TF homolog, using the methods described above. The selected cDNAs may be confirmed by sequencing and enzymatic activity.

3. Altered Expression of Transcription Factors

Any of the identified sequences may be incorporated into a cassette or vector for expression in plants. A number of expression vectors suitable for stable transformation of plant cells or for the establishment of transgenic plants have been described including those described in Weissbach and Weissbach, (1989) *Methods for Plant Molecular Biology*, Academic Press, and Gelvin et al., (1990) *Plant Molecular Biology Manual*, Kluwer Academic Publishers. Specific examples include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, as well as those disclosed by Herrera-Estrella, L., et al., (1983) *Nature* 303: 209, Bevan, M., *Nucl. Acids Res.* (1984) 12: 8711-8721, Klee, H. J., (1985) *Bio/Technology* 3: 637-642, for dicotyledonous plants. Ti-derived plasmids can be transferred into both

monocotyledonous and dicotyledonous species using *Agrobacterium*-mediated transformation (Ishida et al (1996) *Nat. Biotechnol.* 14:745-50; Barton et al. (1983) *Cell* 32:1033-1043).

Alternatively, non-Ti vectors can be used to transfer the DNA into plants and cells by using free DNA delivery techniques. Such methods may involve, for example, the use of liposomes, electroporation, microprojectile bombardment, silicon carbide whiskers, and viruses. By using these methods transgenic plants such as wheat, rice (Christou, P., (1991) *Bio/Technology* 9: 957-962) and corn (Gordon-Kamm, W., (1990) *Plant Cell* 2: 603-618) can be produced. An immature embryo can also be a good target tissue for monocots for direct DNA delivery techniques by using the particle gun (Weeks, T. et al., (1993) *Plant Physiol.* 102: 1077-1084; Vasil, V., (1993) *Bio/Technology* 10: 667-674; Wan, Y. and Lemeaux, P., (1994) *Plant Physiol.* 104: 37-48, and for *Agrobacterium*-mediated DNA transfer (Ishida et al., (1996) *Nature Biotech.* 14: 745-750).

Typically, plant transformation vectors include one or more cloned plant coding sequences (genomic or cDNA) under the transcriptional control of 5' and 3' regulatory sequences and a dominant selectable marker. Such plant transformation vectors typically also contain a promoter (e.g., a regulatory region controlling inducible or constitutive, environmentally-or developmentally-regulated, or cell- or tissue-specific expression), a transcription initiation start site, an RNA processing signal (such as intron splice sites), a transcription termination site, and/or a polyadenylation signal.

Examples of constitutive plant promoters which may be useful for expressing the TF sequence include: the cauliflower mosaic virus (CaMV) 35S promoter, which confers constitutive, high-level expression in most plant tissues (see, e.g., Odel et al., (1985) *Nature* 313:810); the nopaline synthase promoter (An et al., (1988) *Plant Physiol.* 88:547); and the octopine synthase promoter (Fromm et al., (1989) *Plant Cell* 1: 977).

A variety of plant gene promoters that regulate gene expression in response to environmental, hormonal, chemical, developmental signals, and in a tissue-active manner can be used for expression of the TFs in plants, as illustrated by seed-specific promoters (such as the napin, phaseolin or DC3 promoter described in US Pat. No. 5,773,697), root-specific promoters, such as those disclosed in US Patent Nos. 5,618,988, 5,837,848 and 5,905,186; fruit-specific promoters that are active during fruit ripening (such as the *dru 1* promoter (US Pat. No. 5,783,393), or the 2A11 promoter (US Pat. No. 4,943,674) and the tomato polygalacturonase promoter (Bird et al. (1988) *Plant Mol. Biol.* 11:651), root-specific promoters, such as those disclosed in US Patent Nos. 5,618,988, 5,837,848 and 5,905,186, pollen-active promoters such as PTA29, PTA26 and PTA13 (US Pat. No. 5,792,929), promoters active in vascular tissue (Ringli and Keller (1998) *Plant Mol. Biol.* 37:977-988), flower-specific (Kaiser et al, (1995) *Plant Mol. Biol.* 28:231-243), pollen (Baerson et al. (1994) *Plant Mol. Biol.* 26:1947-1959), carpels (Ohl et al. (1990) *Plant Cell* 2:837-848), pollen and

ovules (Baerson et al. (1993) *Plant Mol. Biol.* 22:255-267) auxin-inducible promoters (such as that described in van der Kop et al (1999) *Plant Mol. Biol.* 39:979-990 or Baumann et al. (1999) *Plant Cell* 11:323-334), cytokinin-inducible promoter (Guevara-Garcia (1998) *Plant Mol. Biol.* 38:743-753), promoters responsive to gibberellin (Shi et al. (1998) *Plant Mol. Biol.* 38:1053-1060, Willmott et al. (1998) 38:817-825) and the like. Additional promoters are those that elicit expression in response to heat (Ainley, et al. (1993) *Plant Mol. Biol.* 22: 13-23), light (e.g., the pea *rbcS*-3A promoter, Kuhlemeier et al., (1989) *Plant Cell* 1:471, and the maize *rbcS* promoter, Schaffner and Sheen, (1991) *Plant Cell* 3: 997); wounding (e.g., *wun1*, Siebertz et al., (1989) *Plant Cell* 1: 961); pathogen resistance, and chemicals such as methyl jasmonate or salicylic acid (Gatz et al., (1997) *Plant Mol. Biol.* 48: 89-108). In addition, the timing of the expression can be controlled by using promoters such as those acting at late seed development (Odell et al. (1994) *Plant Physiol.* 106:447-458).

Plant expression vectors may also include RNA processing signals that may be positioned within, upstream or downstream of the coding sequence. In addition, the expression vectors may include additional regulatory sequences from the 3'-untranslated region of plant genes, e.g., a 3' terminator region to increase mRNA stability of the mRNA, such as the PI-II terminator region of potato or the octopine or nopaline synthase 3' terminator regions.

Finally, as noted above, plant expression vectors may also include dominant selectable marker genes to allow for the ready selection of transformants. Such genes include those encoding antibiotic resistance genes (e.g., resistance to hygromycin, kanamycin, bleomycin, G418, streptomycin or spectinomycin) and herbicide resistance genes (e.g., phosphinothricin acetyltransferase).

A reduction of TF expression in a transgenic plant to modify a plant trait may be obtained by introducing into plants antisense constructs based on the TF cDNA. For antisense suppression, the TF cDNA is arranged in reverse orientation relative to the promoter sequence in the expression vector. The introduced sequence need not be the full length TF cDNA or gene, and need not be identical to the TF cDNA or a gene found in the plant type to be transformed. Generally, however, where the introduced sequence is of shorter length, a higher degree of homology to the native TF sequence will be needed for effective antisense suppression. Preferably, the introduced antisense sequence in the vector will be at least 30 nucleotides in length, and improved antisense suppression will typically be observed as the length of the antisense sequence increases. Preferably, the length of the antisense sequence in the vector will be greater than 100 nucleotides. Transcription of an antisense construct as described results in the production of RNA molecules that are the reverse complement of mRNA molecules transcribed from the endogenous TF gene in the plant cell. Suppression of endogenous TF gene expression can also be achieved using a ribozyme. Ribozymes are

synthetic RNA molecules that possess highly specific endoribonuclease activity. The production and use of ribozymes are disclosed in U.S. Patent No. 4,987,071 to Cech and U.S. Patent No. 5,543,508 to Haselhoff. The inclusion of ribozyme sequences within antisense RNAs may be used to confer RNA cleaving activity on the antisense RNA, such that endogenous mRNA molecules that bind to the antisense RNA are cleaved, which in turn leads to an enhanced antisense inhibition of endogenous gene expression.

Vectors in which RNA encoded by the TF cDNA (or variants thereof) is over-expressed may also be used to obtain co-suppression of the endogenous TF gene in the manner described in U.S. Patent No. 5,231,020 to Jorgensen. Such co-suppression (also termed sense suppression) does not require that the entire TF cDNA be introduced into the plant cells, nor does it require that the introduced sequence be exactly identical to the endogenous TF gene. However, as with antisense suppression, the suppressive efficiency will be enhanced as (1) the introduced sequence is lengthened and (2) the sequence similarity between the introduced sequence and the endogenous TF gene is increased.

Vectors expressing an untranslatable form of the TF mRNA may also be used to suppress the expression of endogenous TF activity to modify a trait. Methods for producing such constructs are described in U.S. Patent No. 5,583,021 to Dougherty et al. Preferably, such constructs are made by introducing a premature stop codon into the TF gene. Alternatively, a plant trait may be modified by gene silencing using double-strand RNA (Sharp (1999) *Genes and Development* 13: 139-141).

Another method for abolishing the expression of a gene is by insertion mutagenesis using the T-DNA of *Agrobacterium tumefaciens*. After generating the insertion mutants, the mutants can be screened to identify those containing the insertion in a TF gene. Mutants containing a single mutation event at the desired gene may be crossed to generate homozygous plants for the mutation (Koncz et al. (1992) *Methods in Arabidopsis Research*. World Scientific).

A plant trait may also be modified by using the cre-lox system (for example, as described in US Pat. No. 5,658,772). A plant genome may be modified to include first and second lox sites that are then contacted with a Cre recombinase. If the lox sites are in the same orientation, the intervening DNA sequence between the two sites is excised. If the lox sites are in the opposite orientation, the intervening sequence is inverted.

The polynucleotides and polypeptides of this invention may also be expressed in a plant in the absence of an expression cassette by manipulating the activity or expression level of the endogenous gene by other means. For example, by ectopically expressing a gene by T-DNA activation tagging (Ichikawa et al., (1997) *Nature* 390 698-701, Kakimoto et al., (1996) *Science* 274: 982-985). This method entails transforming a plant with a gene tag containing multiple transcriptional enhancers and once the tag has inserted into the genome, expression of a flanking gene coding sequence becomes deregulated. In another example, the

transcriptional machinery in a plant may be modified so as to increase transcription levels of a polynucleotide of the invention (See PCT Publications WO9606166 and WO 9853057 which describe the modification of the DNA binding specificity of zinc finger proteins by changing particular amino acids in the DNA binding motif).

5 The transgenic plant may also comprise the machinery necessary for expressing or altering the activity of a polypeptide encoded by an endogenous gene, for example by altering the phosphorylation state of the polypeptide to maintain it in an activated state.

4. Transgenic Plants with Modified TF Expression

10 Once an expression cassette comprising a polynucleotide encoding a TF gene of this invention has been constructed, standard techniques may be used to introduce the polynucleotide into a plant in order to modify a trait of the plant. The plant may be any higher plant, including gymnosperms, monocotyledonous and dicotyledonous plants. Suitable protocols are available for *Leguminosae* (alfalfa, soybean, clover, etc.), *Umbelliferae* (carrot, celery, parsnip), *Cruciferae* (cabbage, radish, rapeseed, broccoli, etc.), *Curcubitaceae* (melons and cucumber), *Gramineae* (wheat, corn, rice, barley, millet, etc.), *Solanaceae* (potato, tomato, tobacco, peppers, etc.), and various other crops. See protocols described in Ammirato et al. (1984) *Handbook of Plant Cell Culture –Crop Species*. Macmillan Publ. Co. Shimamoto et al. (1989) *Nature* 338:274-276; Fromm et al. (1990) *Bio/Technology* 8:833-15 839; and Vasil et al. (1990) *Bio/Technology* 8:429-434.

20 Transformation and regeneration of both monocotyledonous and dicotyledonous plant cells is now routine, and the selection of the most appropriate transformation technique will be determined by the practitioner. The choice of method will vary with the type of plant to be transformed; those skilled in the art will recognize the suitability of particular methods for given plant types. Suitable methods may include, but are not limited to: electroporation of plant protoplasts; liposome-mediated transformation; polyethylene glycol (PEG) mediated transformation; transformation using viruses; micro-injection of plant cells; micro-projectile bombardment of plant cells; vacuum infiltration; and *Agrobacterium tumefaciens* mediated transformation. Transformation means introducing a nucleotide sequence in a plant in a 25 manner to cause stable or transient expression of the sequence.

30 Successful examples of the modification of plant characteristics by transformation with cloned sequences which serve to illustrate the current knowledge in this field of technology, and which are herein incorporated by reference, include: U.S. Patent Nos. 5,571,706; 5,677,175; 5,510,471; 5,750,386; 5,597,945; 5,589,615; 5,750,871; 5,268,526; 5,780,708; 35 5,538,880; 5,773,269; 5,736,369 and 5,610,042.

Following transformation, plants are preferably selected using a dominant selectable marker incorporated into the transformation vector. Typically, such a marker will confer

antibiotic or herbicide resistance on the transformed plants, and selection of transformants can be accomplished by exposing the plants to appropriate concentrations of the antibiotic or herbicide.

After transformed plants are selected and grown to maturity, those plants showing a modified trait are identified. The modified trait may be any of those traits described above. Additionally, to confirm that the modified trait is due to changes in expression levels or activity of the polypeptide or polynucleotide of the invention may be determined by analyzing mRNA expression using Northern blots, RT-PCR or microarrays, or protein expression using immunoblots or Western blots or gel shift assays.

The plants may have commercial utility for increasing tolerance or resistance to pathogens and pests. These transgenic plants may be more resistant to biotrophic or necrotrophic pathogens or belonging to the following groups such as a fungus, bacterium, mollicute, virus, nematode, a parasitic higher plant or the like and associated diseases. In particular, pathogens such as *Fusarium oxysporum*, *Erysiphe orontii* and other powdery mildews, *Sclerotinia spp.*, soil-borne oomycetes, foliar oomycetes, *Botrytis spp.*, *Rhizoctonia spp.*, *Verticillium dahliae/albo-atrum*, *Alternaria spp.*, rusts, *Mycosphaerella spp.*, *Fusarium solani*, or the like. The diseases include fungal diseases such as rusts, smuts, wilts, yellows, root rot, leaf drop, ergot, leaf blight of potato, brown spot of rice, leaf blight, late blight, powdery mildew, downy mildew, and the like; viral diseases such as sugarcane mosaic, cassava mosaic, sugar beet yellows, plum pox, barley yellow dwarf, tomato yellow leaf curl, tomato spotted wilt virus, and the like; bacterial diseases such as citrus canker, bacterial leaf blight, bacterial wilt, soft rot of vegetables, and the like; nematode diseases caused by parasitic nematodes such as root-knot nematodes, cyst nematodes or the like.

5. Other Utility of the Polypeptide and Polynucleotide Sequences

A transcription factor provided by the present invention may also be used to identify exogenous or endogenous molecules that may affect expression of the transcription factors and may affect any of the traits described herein. These molecules may include organic or inorganic compounds.

For example, the method may entail first placing the molecule in contact with a plant or plant cell. The molecule may be introduced by topical administration, such as spraying or soaking of a plant, and then the molecule's effect on the expression or activity of the TF polypeptide or the expression of the polynucleotide monitored. Changes in the expression of the TF polypeptide may be monitored by use of polyclonal or monoclonal antibodies, gel electrophoresis or the like. Changes in the expression of the corresponding polynucleotide sequence may be detected by use of microarrays, Northern blots or any other technique for monitoring changes in mRNA expression. These techniques are exemplified in Ausubel et al.

(eds) *Current Protocols in Molecular Biology*, John Wiley & Sons (1998). Such changes in the expression levels may be correlated with modified plant traits and thus identified molecules may be useful for soaking or spraying on fruit, vegetable and grain crops to modify traits in plants.

5 The transcription factors may also be employed to identify promoter sequences with which they may interact. After identifying a promoter sequence, interactions between the transcription factor and the promoter sequence may be modified by changing specific nucleotides in the promoter sequence or specific amino acids in the transcription factor that interact with the promoter sequence to alter a plant trait. Typically, transcription factor DNA
10 binding sites are identified by gel shift assays. After identifying the promoter regions, the promoter region sequences may be employed in double-stranded DNA arrays to identify molecules that affect the interactions of the TFs with their promoters (Bulyk et al. (1999) *Nature Biotechnology* 17:573-577).

15 The identified transcription factors are also useful to identify proteins that modify the activity of the transcription factor. Such modification may occur by covalent modification, such as by phosphorylation, or by protein-protein (homo or-heteropolymer) interactions. Any method suitable for detecting protein-protein interactions may be employed. Among the methods that may be employed are co-immunoprecipitation, cross-linking and co-purification through gradients or chromatographic columns, and the two-hybrid yeast system.

20 The two-hybrid system detects protein interactions in vivo and is described in Chien, et al., (1991), *Proc. Natl. Acad. Sci. USA*, 88, 9578-9582 and is commercially available from Clontech (Palo Alto, Calif.). In such a system, plasmids are constructed that encode two hybrid proteins: one consists of the DNA-binding domain of a transcription activator protein fused to the TF polypeptide and the other consists of the transcription activator protein's
25 activation domain fused to an unknown protein that is encoded by a cDNA that has been recombined into the plasmid as part of a cDNA library. The DNA-binding domain fusion plasmid and the cDNA library are transformed into a strain of the yeast *Saccharomyces cerevisiae* that contains a reporter gene (e.g., lacZ) whose regulatory region contains the transcription activator's binding site. Either hybrid protein alone cannot activate transcription of
30 the reporter gene. Interaction of the two hybrid proteins reconstitutes the functional activator protein and results in expression of the reporter gene, which is detected by an assay for the reporter gene product. Then, the library plasmids responsible for reporter gene expression are isolated and sequenced to identify the proteins encoded by the library plasmids. After identifying proteins that interact with the transcription factors, assays for compounds that
35 interfere with the TF protein-protein interactions may be preformed.

 The following examples are intended to illustrate but not limit the present invention.

Example I. Full Length Gene Identification and Cloning

Putative transcription factor sequences (genomic or ESTs) related to known transcription factors were identified in the *Arabidopsis thaliana* GenBank database using the tblastn sequence analysis program using default parameters and a P-value cutoff threshold of
 5 −4 or −5 or lower, depending on the length of the query sequence. Putative transcription factor sequence hits were then screened to identify those containing particular sequence strings. If the sequence hits contained such sequence strings, the sequences were confirmed as transcription factors.

Alternatively, *Arabidopsis thaliana* cDNA libraries derived from different tissues or
 10 treatments, or genomic libraries were screened to identify novel members of a transcription family using a low stringency hybridization approach. Probes were synthesized using gene specific primers in a standard PCR reaction (annealing temperature 60° C) and labeled with ³²P dCTP using the High Prime DNA Labeling Kit (Boehringer Mannheim). Purified
 15 radiolabelled probes were added to filters immersed in Church hybridization medium (0.5 M NaPO₄ pH 7.0, 7% SDS, 1 % w/v bovine serum albumin) and hybridized overnight at 60 °C with shaking. Filters were washed two times for 45 to 60 minutes with 1xSSC, 1% SDS at 60° C.

To identify additional sequence 5' or 3' of a partial cDNA sequence in a cDNA library, 5' and 3' rapid amplification of cDNA ends (RACE) was performed using the Marathon™
 20 cDNA amplification kit (Clontech, Palo Alto, CA). Generally, the method entailed first isolating poly(A) mRNA, performing first and second strand cDNA synthesis to generate double stranded cDNA, blunting cDNA ends, followed by ligation of the Marathon™ Adaptor to the cDNA to form a library of adaptor-ligated ds cDNA. Gene-specific primers were designed
 25 to be used along with adaptor specific primers for both 5' and 3' RACE reactions. Nested primers, rather than single primers, were used to increase PCR specificity. Using 5' and 3' RACE reactions, 5' and 3' RACE fragments were obtained, sequenced and cloned. The process may be repeated until 5' and 3' ends of the full-length gene were identified. Then the full-length cDNA was generated by PCR using primers specific to 5' and 3' ends of the gene by end-to-end PCR.

Example II Pathogen Resistance Genes

RT-PCR and microarray experiments were performed to identify those genes induced after exposure to biotrophic fungal pathogens, such as *Erisyphe orontii*, necrotropic fungal
 35 pathogens, such as *Fusarium oxysporum*, and disease associated growth-regulators such as salicylic acid, methyl jasmonate and ethylene (ACC). The gene expression patterns from soil grown as well as tissue culture grown plant tissue were investigated.

Fusarium oxysporum isolates cause vascular wilts and damping off of various annual vegetables, perennials and weeds (Mauch-Mani and Slusarenko (1994) Molecular Plant-Microbe Interactions 7: 378-383). For *Fusarium oxysporum* experiments, plants grown on petri dishes were sprayed with a fresh spore suspension of *F. oxysporum*. The spore suspension was prepared as follows: A plug of fungal hyphae from a plate culture was placed on a fresh potato dextrose agar plate and allowed to spread for one week. 5 ml sterile water was then added to the plate, swirled, and pipetted into 50 ml Armstrong Fusarium medium. Spores were grown overnight in Fusarium medium and then sprayed onto plants using a Preval paint sprayer. Plant tissue was harvested and frozen in liquid nitrogen 48 hours post infection

Erysiphe orontii is a causal agent of powdery mildew. For *Erysiphe orontii* experiments, plants were grown approximately 4 weeks in a greenhouse under 12 hour light (20 C, ~30% relative humidity (rh)). Individual leaves were infected with *E. orontii* spores from infected plants using a camel's hair brush, and the plants were transferred to a Percival growth chamber (20 C, 80% rh.). Plant tissue was harvested and frozen in liquid nitrogen 7 days post infection.

For salicylic acid experiments, 15 day old seedlings grown on petri dishes were transferred to plates containing 0.5 mM salicylic acid (SA). After 72 hours, leaves were harvested and frozen in liquid nitrogen.

Reverse transcriptase PCR was done using gene specific primers within the coding region for each sequence identified. The primers were designed near the 3' region of each coding sequence initially identified.

Total RNA from these tissues were isolated using the CTAB extraction protocol. Once extracted total RNA was normalized in concentration across all the tissue types to ensure that the PCR reaction for each tissue received the same amount of cDNA template using the 28S band as reference. Poly A+ was purified using a modified protocol from the Qiagen Oligotex kit batch protocol. cDNA was synthesized using standard protocols. After the first strand cDNA synthesis, primers for Actin 2 were used to normalize the concentration of cDNA across the tissue types. Actin 2 is found to be constitutively expressed in fairly equal levels across the tissue types we are investigating.

For RT PCR, cDNA template was mixed with corresponding primers and Taq polymerase. Each reaction consisted of 0.2 ul cDNA template, 2ul 10X Tricine buffer, 2 ul 10X Tricine buffer and 16.8 ul water, 0.05ul Primer 1, 0.05 ul, Primer 2, 0.3 ul Taq polymerase and 8.6 ul water.

The 96 well plate was covered with microfilm and set in the Thermocycler to start the following reaction cycle. Step1 93° C for 3 mins, Step 2 93° C for 30 sec, Step 3 65° C for 1 min, Step 4 72° C for 2 mins,. Steps 2, 3 and 4 were repeated for 28 cycles, Step 5 72° C

for 5 mins and Step 6 4° C. The PCR plate was placed back in the thermocycler to amplify more products at 8 more cycles to identify genes that have very low expression. The reaction cycle was as follows: Step 2 93° C for 30 sec, Step 3 65° C for 1 min, and Step 4 72° C for 2 mins, repeated for 8 cycles, and Step 4 4° C.

8ul of PCR product and 1.5 ul of loading dye were loaded on a 1.2% agarose gel for analysis after 28 cycles and 36 cycles. Expression levels of specific transcripts were considered low if they were only detectable after 36 cycles of PCR. Expression levels were considered medium or high depending on the levels of transcript compared with observed transcript levels for actin2.

In some instances, expression patterns of the transcription factors was monitored by microarray experiments. cDNAs were generated by PCR and resuspended at a final concentration of ~ 100 ng/ul in 3X SSC or 150mM Na-phosphate (Eisen and Brown (1999) *Meth. in Enzymol.* 303:179-205). The cDNAs were spotted on microscope glass slides coated with polylysine. The prepared cDNAs were aliquoted into 384 well plates and spotted on the slides using an x-y-z gantry (OmniGrid) purchased from GeneMachines (Menlo Park, CA) outfitted with quill type pins purchased from Telechem International (Sunnyvale, CA). After spotting, the arrays were cured for a minimum of one week at room temperature, rehydrated and blocked following the protocol recommended by Eisen and Brown (1999).

Sample total RNA (10 ug) samples were labeled using fluorescent Cy3 and Cy5 dyes. Labeled samples were resuspended in 4X SSC/0.03% SDS/4 ug salmon sperm DNA/2 ug tRNA/ 50mM Na-pyrophosphate, heated for 95°C for 2.5 minutes, spun down and placed on the array. The array was then covered with a glass coverslip and placed in a sealed chamber. The chamber was then kept in a water bath at 62°C overnight. The arrays were washed as described in Eisen and Brown (1999) and scanned on a General Scanning 3000 laser scanner. The resulting files are subsequently quantified using Imagen software purchased from BioDiscovery (Los Angeles, CA).

The transcript levels were observed to be upregulated between 1.5 and 100 fold when compared with control plants not exposed to the pathogens.

Example III. Construction of Expression Vectors

The sequence was amplified from a genomic or cDNA library using primers specific to sequences upstream and downstream of the coding region. The expression vector was pMEN20, which is derived from pMON316 (Sanders et al, (1987) *Nucleic Acids Research* 15:1543-58). To clone the sequence into the vector, both pMEN20 and the amplified DNA fragment were digested separately with Sall and NotI restriction enzymes at 37° C for 2 hours. The digestion products were subject to electrophoresis in a 0.8% agarose gel and visualized

by ethidium bromide staining. The DNA fragments containing the sequence and the linearized plasmid were excised and purified by using a Qiaquick gel extraction kit (Qiagen, CA). The fragments of interest were ligated at a ratio of 3:1 (vector to insert). Ligation reactions using T4 DNA ligase (New England Biolabs, MA) were carried out at 16° C for 16 hours. The ligated DNAs were transformed into competent cells of the *E. coli* strain DH5alpha by using the heat shock method. The transformations were plated on LB plates containing 50 mg/l spectinomycin (Sigma).

Individual colonies were grown overnight in five milliliters of LB broth containing 50 mg/l spectinomycin at 37° C. Plasmid DNA was purified by using Qiaquick Mini Prep kits (Qiagen, CA).

Example IV. Transformation of *Agrobacterium* with the Expression Vector

After the plasmid vector containing the gene was constructed, the vector was used to transform *Agrobacterium tumefaciens* cells expressing the gene products. The stock of *Agrobacterium tumefaciens* cells for transformation were made as described by Nagel et al. *FEMS Microbiol Letts* 67: 325-328 (1990). *Agrobacterium* strain GV3101 was grown in 250 ml LB medium (Sigma) overnight at 28°C with shaking until an absorbance (A_{600}) of 0.5 – 1.0 was reached. Cells were harvested by centrifugation at 4,000 x g for 15 min at 4° C. Cells were then resuspended in 250 µl chilled buffer (1 mM HEPES, pH adjusted to 7.0 with KOH). Cells were centrifuged again as described above and resuspended in 125 µl chilled buffer. Cells were then centrifuged and resuspended two more times in the same HEPES buffer as described above at a volume of 100 µl and 750 µl, respectively. Resuspended cells were then distributed into 40 µl aliquots, quickly frozen in liquid nitrogen, and stored at -80° C.

Agrobacterium cells were transformed with plasmids prepared as described above following the protocol described by Nagel et al. *FEMS Microbiol Letts* 67: 325-328 (1990). For each DNA construct to be transformed, 50 – 100 ng DNA (generally resuspended in 10 mM Tris-HCl, 1 mM EDTA, pH 8.0) was mixed with 40 µl of *Agrobacterium* cells. The DNA/cell mixture was then transferred to a chilled cuvette with a 2mm electrode gap and subject to a 2.5 kV charge dissipated at 25 µF and 200 µF using a Gene Pulser II apparatus (Bio-Rad). After electroporation, cells were immediately resuspended in 1.0 ml LB and allowed to recover without antibiotic selection for 2 – 4 hours at 28° C in a shaking incubator. After recovery, cells were plated onto selective medium of LB broth containing 100 µg/ml spectinomycin (Sigma) and incubated for 24-48 hours at 28° C. Single colonies were then picked and inoculated in fresh medium. The presence of the plasmid construct was verified by PCR amplification and sequence analysis.

Example V. Transformation of *Arabidopsis* Plants with *Agrobacterium tumefaciens* with Expression Vector

After transformation of *Agrobacterium tumefaciens* with plasmid vectors containing the gene, single *Agrobacterium* colonies were identified, propagated, and used to transform *Arabidopsis* plants. Briefly, 500 ml cultures of LB medium containing 50 mg/l spectinomycin were inoculated with the colonies and grown at 28° C with shaking for 2 days until an absorbance (A_{600}) of > 2.0 is reached. Cells were then harvested by centrifugation at 4,000 x g for 10 min, and resuspended in infiltration medium (1/2 X Murashige and Skoog salts (Sigma), 1 X Gamborg's B-5 vitamins (Sigma), 5.0% (w/v) sucrose (Sigma), 0.044 μ M benzylamino purine (Sigma), 200 μ L/L Silwet L-77 (Lehle Seeds) until an absorbance (A_{600}) of 0.8 was reached.

Prior to transformation, *Arabidopsis thaliana* seeds (ecotype Columbia) were sown at a density of ~10 plants per 4" pot onto Pro-Mix BX potting medium (Hummert International) covered with fiberglass mesh (18 mm X 16 mm). Plants were grown under continuous illumination (50-75 μ E/m²/sec) at 22-23° C with 65-70% relative humidity. After about 4 weeks, primary inflorescence stems (bolts) are cut off to encourage growth of multiple secondary bolts. After flowering of the mature secondary bolts, plants were prepared for transformation by removal of all siliques and opened flowers.

The pots were then immersed upside down in the mixture of *Agrobacterium* infiltration medium as described above for 30 sec, and placed on their sides to allow draining into a 1' x 2' flat surface covered with plastic wrap. After 24 h, the plastic wrap was removed and pots are turned upright. The immersion procedure was repeated one week later, for a total of two immersions per pot. Seeds were then collected from each transformation pot and analyzed following the protocol described below.

Example VI. Identification of *Arabidopsis* Primary Transformants

Seeds collected from the transformation pots were sterilized essentially as follows. Seeds were dispersed into in a solution containing 0.1% (v/v) Triton X-100 (Sigma) and sterile H₂O and washed by shaking the suspension for 20 min. The wash solution was then drained and replaced with fresh wash solution to wash the seeds for 20 min with shaking. After removal of the second wash solution, a solution containing 0.1% (v/v) Triton X-100 and 70% ethanol (Equistar) was added to the seeds and the suspension was shaken for 5 min. After removal of the ethanol/detergent solution, a solution containing 0.1% (v/v) Triton X-100 and 30% (v/v) bleach (Clorox) was added to the seeds, and the suspension was shaken for 10 min. After removal of the bleach/detergent solution, seeds were then washed five times in sterile distilled H₂O. The seeds were stored in the last wash water at 4° C for 2 days in the dark before being plated onto antibiotic selection medium (1 X Murashige and Skoog salts (pH

adjusted to 5.7 with 1M KOH), 1 X Gamborg's B-5 vitamins, 0.9% phytagar (Life Technologies), and 50 mg/l kanamycin). Seeds were germinated under continuous illumination (50-75 $\mu\text{E}/\text{m}^2/\text{sec}$) at 22-23°C. After 7-10 days of growth under these conditions, kanamycin resistant primary transformants (T_1 generation) were visible and obtained. These seedlings were transferred first to fresh selection plates where the seedlings continued to grow for 3-5 more days, and then to soil (Pro-Mix BX potting medium).

Primary transformants are self-crossed and progeny seeds (T_2) collected.

Example VII. Analysis of Arabidopsis T_2 progeny plants for Pathogen Resistance or Pathogen Tolerance

T_2 or knockout mutant seed were surface sterilized and sown on MS media containing sucrose. Ten days post-planting, seedlings were transferred to MS media without sucrose. At two weeks of age *Arabidopsis* seedlings were inoculated with *Fusarium* by spraying with a spore suspension (2×10^6 conidia per milliliter) and incubated under high humidity. Plants were then scored macroscopically for disease symptoms or microscopically for fungal growth or using microarrays for the induction of resistance associated genes (such as the defensin genes) to detect resistance or tolerance of the plant tissue. A wild type plant shows the first signs of damage (gradual yellowing of leaves, damping off of seedlings or growth of fungal mycelium) after two to four days after inoculation. Transgenic plants that are pathogen resistant or tolerant showed a delay in disease or symptom development compared to wild-type control plants.

Alternatively, *Erysiphe* inoculations were done by tapping conidia from 1 to 2 heavily infected leaves onto the mesh cover of a settling tower, brushing the mesh with a camel's hair paint brush to break up the conidial chains, and letting the conidia settle for 10 minutes. Plants were 4 to 4.5 weeks old at the time of inoculation. Spores were obtained from 10 to 14 day old *Erysiphe* cultures. Typically, within the first twenty-four hours, the spores differentiated into several fungal structures including the haustorium that invaginates a host's epidermal plasma membrane. Formation of aerial mycelium and sporulation represent late differentiation events between 4 and 7 days post inoculation (Freilaldenhoven et al. (1994) *Plant Cell* 6: 983-994). Plant resistance was scored based on the relative number and size of mycelial patches bearing conidia compared to wild-type control plants. Events associated with disease resistance to the pathogens and pests include: the induction of pathogen resistance related genes (R genes), the activation of cell death in the attacked epidermal cells (hypersensitive response), the induction of anti-microbial compounds, such as phytoalexins, and the lignification that occurs at attempted penetration sites. Assays are performed to observe these events. Transgenic plants identified that induce R genes, activate cell death, induce anti-microbial compounds or increase lignification sooner or to a greater extent than

wild-type plants when exposed to pathogen are potentially more resistant to infection by *Erysiphe* as well as a number of other pathogens and pests.

We have observed that when the expression levels of the genes are altered, that the disease phenotype can be varied. For example, G19 was significantly induced upon infection by the fungal pathogen *Erysiphe orontii* as well as the disease associated growth regulator, ethylene. Our data show that G19 overexpressing plants were more tolerant to infection with a moderate dose of *Erysiphe orontii* and in a nematode screen. The transgenic plants overexpressing G19 under the control of the 35S promoter were morphologically similar to control plants.

Additionally, G511 was another example of a gene that when overexpressed showed an increased tolerance to the fungal pathogen *Erysiphe orontii*. In both cases increased tolerance includes a significant reduction in pathogen growth and symptom development compared to wild type plants that were treated with pathogen in an identical manner.

Example VIII. Transformation of Cereal Plants with the Expression Vector

A cereal plant, such as corn, wheat, rice, sorghum or barley, can also be transformed with the plasmid vectors containing the sequence and constitutive or inducible promoters to modify a trait. In these cases, a cloning vector, pMEN020, is modified to replace the NptII coding region with the BAR gene of *Streptomyces hygroscopicus* that confers resistance to phosphinothricin. The KpnI and BglII sites of the Bar gene are removed by site-directed mutagenesis with silent codon changes.

Plasmids according to the present invention may be transformed into corn embryogenic cells derived from immature scutellar tissue by using microprojectile bombardment, with the A188XB73 genotype as the preferred genotype (Fromm et al., *Bio/Technology* 8: 833-839 (1990); Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990)). After microprojectile bombardment the tissues are selected on phosphinothricin to identify the transgenic embryogenic cells (Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990)). Transgenic plants are regenerated by standard corn regeneration techniques (Fromm, et al., *Bio/Technology* 8: 833-839 (1990); Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990)).

Example IX. Identification of Homologous Sequences

Homologs from the same plant, different plant species or other organisms were identified using database sequence search tools, such as the Basic Local Alignment Search Tool (BLAST) (Altschul et al. (1990) *J. Mol. Biol.* 215:403-410; and Altschul et al. (1997) *Nucl. Acid Res.* 25: 3389-3402). The tblastn or blastn sequence analysis programs were employed using the BLOSUM-62 scoring matrix (Henikoff, S. and Henikoff, J. G. (1992) *Proc. Natl. Acad. Sci. USA* 89: 10915-10919). The output of a BLAST report provides a score that takes

into account the alignment of similar or identical residues and any gaps needed in order to align the sequences. The scoring matrix assigns a score for aligning any possible pair of sequences. The P values reflect how many times one expects to see a score occur by chance. Higher scores are preferred and a low threshold P value threshold is preferred.

These are the sequence identity criteria. The tblastn sequence analysis program was used to query a polypeptide sequence against six-way translations of sequences in a nucleotide database. Hits with a P value less than -25, preferably less than -70, and more preferably less than -100, were identified as homologous sequences (exemplary selected sequence criteria). The blastn sequence analysis program was used to query a nucleotide sequence against a nucleotide sequence database. In this case too, higher scores were preferred and a preferred threshold P value was less than -13, preferably less than -50, and more preferably less than -100.

Alternatively, a fragment of a sequence from Figure 1 is ³²P-radiolabeled by random priming (Sambrook et al., (1989) *Molecular Cloning. A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory Press, New York) and used to screen a plant genomic library (the exemplary test polynucleotides). As an example, total plant DNA from *Arabidopsis thaliana*, *Nicotiana tabacum*, *Lycopersicon pimpinellifolium*, *Prunus avium*, *Prunus cerasus*, *Cucumis sativus*, or *Oryza sativa* are isolated according to Stockinger al (Stockinger, E. J., et al., (1996), *J. Heredity*, 87:214-218). Approximately 2 to 10 µg of each DNA sample are restriction digested, transferred to nylon membrane (Micron Separations, Westboro, MA) and hybridized. Hybridization conditions are: 42° C in 50% formamide, 5X SSC, 20 mM phosphate buffer 1X Denhardt's, 10% dextran sulfate, and 100µg/ml herring sperm DNA. Four low stringency washes at RT in 2X SSC, 0.05% sodium sarcosyl and 0.02% sodium pyrophosphate are performed prior to high stringency washes at 55° C in 0.2X SSC, 0.05% sodium sarcosyl and 0.01% sodium pyrophosphate. High stringency washes are performed until no counts are detected in the washout according to Walling et al. (Walling, L. L., et al., (1988) *Nucl. Acids Res.* 16:10477-10492).

All references (publications and patents) are incorporated herein by reference in their entirety for all purposes.

Although the invention has been described with reference to the embodiments and examples above, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims.

We Claim:

1. A transgenic plant comprising a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of SEQ ID Nos. 2N, where N=1-56, wherein the recombinant polynucleotide alters the plant's disease tolerance or resistance when compared with the same trait of another plant lacking the recombinant polynucleotide.

2. The transgenic plant of claim 1, wherein the nucleotide sequence encodes a polypeptide comprising a conserved domain selected from the group consisting of SEQ ID Nos. 2N, where N=1-56

3. The transgenic plant of claim 1, wherein the recombinant polynucleotide further comprises a promoter operably linked to said nucleotide sequence.

4. The transgenic plant of claim 3, wherein said promoter is constitutive or inducible or tissue-active.

5. A method for altering the disease tolerance or resistance of a plant, said method comprising (a) transforming a plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of SEQ ID Nos. 2N, where N=1-56, (b) selecting said transformed plants; and (c) identifying a transformed plant having an altered disease tolerance or resistance.

6. The method of claim 5, wherein the nucleotide sequence encodes a polypeptide comprising a conserved domain selected from the group consisting of SEQ ID Nos. 2N, where N=1-56.

8. The method of claim 5, wherein the recombinant polynucleotide further comprises a promoter operably linked to said nucleotide sequence.

9. The method of claim 8, wherein said promoter is constitutive or inducible or tissue-active.

10. A method for altering the expression levels of at least one gene in a plant, said method comprising (a) transforming the plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of SEQ ID Nos. 2N, where N=1-56; and (b) selecting said transformed plant.

11. The method of claim 10, wherein said recombinant polynucleotide encodes a polypeptide comprising a conserved domain selected from the group consisting of SEQ ID Nos. 2N, where N=1-56.

12. The method of claim 10, wherein the nucleotide sequence further comprises a promoter operably linked to said nucleotide sequence.

13. The method of claim 10, wherein said promoter is constitutive or inducible or tissue-active.

14. A method for altering the disease tolerance or resistance in a plant, said method comprising (a) transforming the plant with a recombinant polynucleotide comprising at least 18 consecutive nucleotides of a sequence selected from the group consisting of SEQ ID Nos. 2N-1, where N= 1-56, and SEQ ID Nos. 113-121; and (b) selecting said transformed plant.

15. A method for altering a plant's trait, said method comprising (a) providing a database sequence; (b) comparing said database sequence with a polypeptide selected from SEQ ID Nos. 2N, where N= 1-56; (c) selecting a database sequence that meets selected sequence criteria; and (d) transforming said selected database sequence in the plant.

16. A method for altering a plant's trait, said method comprising (a) providing a database sequence; (b) comparing said database sequence with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121; (c) selecting a database sequence that meets selected sequence criteria; and (d) transforming said selected database sequence in the plant.

17. A method for altering a plant's trait, said method comprising (a) providing a test polynucleotide; (b) hybridizing said test polynucleotide with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121 at low stringency; and (c) transforming said hybridizing test polynucleotide in a plant to alter a trait of the plant.

ABSTRACT OF THE INVENTION

Recombinant polynucleotides and methods for altering the regulation of gene expression in plants are provided to modify a plant's traits, in particular disease tolerance.

Figure 1a

SEQ ID No	GID No.	Family	Fragment(s)	DNA or protein	coding sequence	conserved domain
1	G1043	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	43-927	
2	G1043	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		120-179
3	G759	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	8-961	
4	G759	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		17-159
5	G185	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	77-988	
6	G185	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		113-172
7	G629	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	169-1275	
8	G629	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		92-152
9	G435	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	32-502	
10	G435	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		4-67
11	G4	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	90-1217	
12	G4	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		121-188
13	G1035	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	103-624	
14	G1035	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		39-91
15	G179	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	68-511	
16	G179	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		65-121
17	G28	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	63-869	
18	G28	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		145-213
19	G1241	MISC	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	108-605	
20	G1241	MISC	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		
21	G19	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	70-816	
22	G19	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		76-145
23	G503	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	80-886	
24	G503	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		12-158
25	G263	HS	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	48-902	
26	G263	HS	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		15-105
27	G921	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	116-1024	
28	G921	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		146-203
29	G1275	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	58-579	
30	G1275	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		113-169

Figure 1b

SEQ ID No	GID No.	Family	Fragments	DNA or protein	coding sequence	conserved domain
31	G242	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	66-983	
32	G242	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		6-105
33	G1006	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	52-783	
34	G1006	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		114-182
35	G1049	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	29-550	
36	G1049	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		77-132
37	G502	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	224-1186	
38	G502	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		10-155
39	G239	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-822	
40	G239	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		21-125
41	G555	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	250-1242	
42	G555	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		38-110
43	G352	Z	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	80-817	
44	G352	Z	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		99-119, 166-186
45	G1352	Z	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	79-900	
46	G1352	Z	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		108-129, 167-188
47	G1089	bZIPt2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	31-2427	
48	G1089	bZIPt2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		425-500
49	G553	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	82-1236	
50	G553	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		94-160
51	G1221	MISC	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	287-2314	
52	G1221	MISC	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		490-515
53	G580	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	43-747	
54	G580	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		162-218
55	G270	AKR	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	43-1350	
56	G270	AKR	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		
57	G201	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-1011	
58	G201	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-114
59	G1417	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	32-1501	
60	G1417	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		239-296

Figure 1c

SEQ ID No	GID No.	Family	Fragsments	DNA or protein	coding sequence	conserved domain
61	G233	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	46-867	
62	G233	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-114
63	G920	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	114-1154	
64	G920	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		152-211
65	G867	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	64-1098	
66	G867	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		59-124
67	G659	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-984	
68	G659	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		16-116
69	G620	CAAT	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	40-666	
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71	G596	AT-Hook	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	168-1121	
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73	G511	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	31-738	
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75	G471	IAA/ARF	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	115-2112	
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77	G385	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	37-2202	
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81	G25	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	80-595	
82	G25	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		47-114
83	G610	BPF-1	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	137-2059	
84	G610	BPF-1	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		577-609
85	G229	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	41-1156	
86	G229	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-120
87	G221	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	115-795	
88	G221	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		21-125
89	G186	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	100-1761	
90	G186	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		312-369

Figure 1d

SEQ ID No	GID No.	Family	Fragsments	DNA or protein	coding sequence	conserved domain
91	G562	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	137-1285	
92	G562	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		253-315
93	G255	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	30-839	
94	G255	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-115
95	G3	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	16-477	
96	G3	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		11-95
97	G713	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	58-765	
98	G713	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		23-86
99	G515	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	154-1170	
100	G515	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		6-144
101	G390	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-2526	
102	G390	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		18-81
103	G1034	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	214-1443	
104	G1034	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		97-160
105	G1149	PAZ	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-2910	
106	G1149	PAZ	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		870-880
107	G1334	CAAT	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	76-885	
108	G1334	CAAT	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		18-190
109	G1650	HLH/MYC	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	84-1199	
110	G1650	HLH/MYC	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		284-334
111	G241	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
112	G241	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-116
113	G348	GATA Zn	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
114	G171	MADS	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
115	G521	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
116	G1274	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
117	G182	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
118	G1290	AKR	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
119	G374	Z	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
120	G682	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		

Figure 1e

SEQ ID No	GID No.	Family	Fragment	DNA or protein	coding sequence	conserved domain
121	G501	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		

As a below-named inventor, I hereby declare that:

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

the specification of which is attached hereto.

I acknowledge the duty to disclose all information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56(a) which states in relevant part: "Each individual associated with the filing and prosecution of a patent application has a duty of candor and good faith in dealing with the Office, which includes a duty to disclose to the Office all information known to that individual to be material to patentability as defined in this section. The duty to disclose all information known to be material to patentability is deemed to be satisfied if all information known to be material to patentability of any claim issued in a patent was cited by the Office or submitted to the Office in the manner prescribed by §§ 1.97(b)-(d) and 1.98.

I hereby claim foreign priority benefits under Title 35 United States Code, § 119(a)-(d) or 365(a)-(b) of any foreign applications for patent or inventor's certificate as indicated below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

I hereby claim the benefit of priority under Title 35 United States Code, § 119(e) of any United States provisional application(s) listed below:

Filing Date:

3/23/99

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Title 18, United States Code, §1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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- the Assignment recorded on

at reel __, frames __ - __.


Pursuant to 37 C.F.R. § 3.73(b) the undersigned Assignee hereby states that evidentiary documents have been reviewed and hereby certifies that, to the best of ASSIGNEE's knowledge and belief, title is in the identified ASSIGNEE.

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Name: Guo-Liang Yu

Title: Senior Vice-President, Research and Development

Date: 03/20/2000

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 Glu Arg Phe Phe Gln Trp Ile Gly Gly Phe Arg Pro Ser Glu Leu Leu
 225 230 235 240
 Asn Val Val Met Pro Tyr Leu Gln Pro Leu Thr Asp Gln Gln Ile Leu
 245 250 255
 Glu Val Arg Asn Leu Gln Gln Ser Ser Gln Gln Ala Glu Asp Ala Leu
 260 265 270
 Ser Gln Gly Ile Asp Lys Leu Gln Gln Ser Leu Ala Glu Ser Ile Val
 275 280 285
 Ile Asp Ala Val Ile Glu Ser Thr His Tyr Pro Thr His Met Ala Ala
 290 295 300
 Ala Ile Glu Asn Leu Gln Ala Leu Glu Gly Phe Val Asn Gln Ala Asp
 305 310 315 320
 His Leu Arg Gln Gln Thr Leu Gln Gln Met Ala Lys Ile Leu Thr Thr
 325 330 335
 Arg Gln Ser Ala Arg Gly Leu Leu Ala Leu Gly Glu Tyr Leu His Arg
 340 345 350
 Leu Arg Ala Leu Ser Ser Leu Trp Ala Ala Arg Pro Gln Glu Pro Thr
 355 360 365

<210> 9
 <211> 627
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G435

<400> 9
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 caagaagctt gagccagatc tgaaacttca actgtcgaac cagcttggtc tacctcaaag 180
 acaagtcgct gtctggttcc aaaacaagcg agccagggtc aagactcagt ctcttgaggt 240
 ccaacactgc actcttcagt ccaagcacga agcagctctc tccgacaagg caaagttaga 300
 gcatcaagtg cagtttctcc aagatgagct gaagagagca aggaatcagc ttgctctggt 360
 cacaaatcaa gattctcctg ttgataattc taatcttggt tcttgatgatg aagatcatga 420
 tgatcaagtg gtggtattcg acgagcttta cgcttgcttt gttagcaatg gacatggatc 480
 ttcatacaacc tcatgggtct gattctgttt cgacgcagac aagattccaa tatatatagt 540
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 taattaaagt cattcagaca ttcacta 627

<210> 10
 <211> 156
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G435

<400> 10
 Met Glu Asn Ser Gln Ser Gln Gly Lys Asn Lys Lys Lys Arg Leu Thr
 1 5 10 15
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 20 25 30
 Leu Glu Pro Asp Leu Lys Leu Gln Leu Ser Asn Gln Leu Gly Leu Pro
 35 40 45
 Gln Arg Gln Val Ala Val Trp Phe Gln Asn Lys Arg Ala Arg Phe Lys
 50 55 60
 Thr Gln Ser Leu Glu Val Gln His Cys Thr Leu Gln Ser Lys His Glu
 65 70 75 80
 Ala Ala Leu Ser Asp Lys Ala Lys Leu Glu His Gln Val Gln Phe Leu
 85 90 95
 Gln Asp Glu Leu Lys Arg Ala Arg Asn Gln Leu Ala Leu Phe Thr Asn
 100 105 110
 Gln Asp Ser Pro Val Asp Asn Ser Asn Leu Gly Ser Cys Asp Glu Asp
 115 120 125
 His Asp Asp Gln Val Val Val Phe Asp Glu Leu Tyr Ala Cys Phe Val
 130 135 140
 Ser Asn Gly His Gly Ser Ser Ser Thr Ser Trp Val
 145 150 155

<210> 11
 <211> 1577
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G4

<400> 11
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 aaaaagagta gagctttcgt gaagccacca tgtgtggagg agctataatc tccgatttca 120
 tacctccgcc gaggtccctc cgcgtcacta acgagtttat ctggccggat ctgaaaaaca 180
 aagtgaagc ttcaaagaag agatcgaata agcgatccga tttcttcgat cttgacgatg 240
 atttcgaagc tgatttccaa gggtttaagg atgactcggc ttttgactgc gaagacgatg 300
 atgatgtctt cgtcaatgtt aagcctttcg tcttcaccgc aactactaag cccgtagctt 360
 ccgctttcgt ctccactgta ggttcagcat atgccaaaga aactgtagag tccgctgagc 420

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aagctgagaa atcttctaag aggaagagga agaatcaata ccgaggggatt aggcagcgtc 480
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gaacattcga cactgctgag gaagcagcaa gagcttatga tgctgcagca cgcagaatcc 600
gtggcacgaa agctaagggtg aattttcccg aggagaagaa ccctagcgtc gtatcccaga 660
aacgtcctag tgctaagact aataatcttc agaaatcagt ggctaaacca aacaaaagcg 720
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ccgagatctc ttcaatgctt gtcaataaca acgaagcatc atttggtgaa gaaaccaatg 1020
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<210> 12
<211> 375
<212> PRT
<213> Arabidopsis thaliana

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<220>
<223> G4

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<400> 12
Met Cys Gly Gly Ala Ile Ile Ser Asp Phe Ile Pro Pro Pro Arg Ser
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Leu Arg Val Thr Asn Glu Phe Ile Trp Pro Asp Leu Lys Asn Lys Val
      20              25              30

Lys Ala Ser Lys Lys Arg Ser Asn Lys Arg Ser Asp Phe Phe Asp Leu
      35              40              45

Asp Asp Asp Phe Glu Ala Asp Phe Gln Gly Phe Lys Asp Asp Ser Ala
      50              55              60

Phe Asp Cys Glu Asp Asp Asp Asp Val Phe Val Asn Val Lys Pro Phe
      65              70              75              80

Val Phe Thr Ala Thr Thr Lys Pro Val Ala Ser Ala Phe Val Ser Thr
      85              90              95

Val Gly Ser Ala Tyr Ala Lys Lys Thr Val Glu Ser Ala Glu Gln Ala
      100             105             110

Glu Lys Ser Ser Lys Arg Lys Arg Lys Asn Gln Tyr Arg Gly Ile Arg
      115             120             125

Gln Arg Pro Trp Gly Lys Trp Ala Ala Glu Ile Arg Asp Pro Arg Lys
      130             135             140

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Gly Ser Arg Glu Trp Leu Gly Thr Phe Asp Thr Ala Glu Glu Ala Ala
 145 150 155 160
 Arg Ala Tyr Asp Ala Ala Ala Arg Arg Ile Arg Gly Thr Lys Ala Lys
 165 170 175
 Val Asn Phe Pro Glu Glu Lys Asn Pro Ser Val Val Ser Gln Lys Arg
 180 185 190
 Pro Ser Ala Lys Thr Asn Asn Leu Gln Lys Ser Val Ala Lys Pro Asn
 195 200 205
 Lys Ser Val Thr Leu Val Gln Gln Pro Thr His Leu Ser Gln Gln Tyr
 210 215 220
 Cys Asn Asn Ser Phe Asp Asn Ser Phe Gly Asp Met Ser Phe Met Glu
 225 230 235 240
 Glu Lys Pro Gln Met Tyr Asn Asn Gln Phe Gly Leu Thr Asn Ser Phe
 245 250 255
 Asp Ala Gly Gly Asn Asn Gly Tyr Gln Tyr Phe Ser Ser Asp Gln Gly
 260 265 270
 Ser Asn Ser Phe Asp Cys Ser Glu Phe Gly Trp Ser Asp His Gly Pro
 275 280 285
 Lys Thr Pro Glu Ile Ser Ser Met Leu Val Asn Asn Asn Glu Ala Ser
 290 295 300
 Phe Val Glu Glu Thr Asn Ala Ala Lys Lys Leu Lys Pro Asn Ser Asp
 305 310 315 320
 Glu Ser Asp Asp Leu Met Ala Tyr Leu Asp Asn Ala Leu Trp Asp Thr
 325 330 335
 Pro Leu Glu Val Glu Ala Met Leu Gly Ala Asp Ala Gly Ala Val Thr
 340 345 350
 Gln Glu Glu Glu Asn Pro Val Glu Leu Trp Ser Leu Asp Glu Ile Asn
 355 360 365
 Phe Met Leu Glu Gly Asp Phe
 370 375

<210> 13
 <211> 903
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G1035

<400> 13
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 atgggatctt ccacaagtgg aaattgctcg tcggtttcaa ccactgggtt agctaactcc 180


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<210> 14
<211> 173
<212> PRT
<213> Arabidopsis thaliana
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<400> 14
Met Asn Asn Lys Thr Glu Met Gly Ser Ser Thr Ser Gly Asn Cys Ser
  1          5          10          15

Ser Val Ser Thr Thr Gly Leu Ala Asn Ser Gly Ser Glu Ser Asp Leu
          20          25          30

Arg Gln Arg Asp Leu Ile Asp Glu Arg Lys Arg Lys Arg Lys Gln Ser
  35          40          45

Asn Arg Glu Ser Ala Arg Arg Ser Arg Met Arg Lys Gln Lys His Leu
  50          55          60

Asp Asp Leu Thr Ala Gln Val Thr His Leu Arg Lys Glu Asn Ala Gln
  65          70          75          80

Ile Val Ala Gly Ile Ala Val Thr Thr Gln His Tyr Val Thr Ile Glu
          85          90          95

Ala Glu Asn Asp Ile Leu Arg Ala Gln Val Leu Glu Leu Asn His Arg
          100          105          110

Leu Gln Ser Leu Asn Glu Ile Val Asp Phe Val Glu Ser Ser Ser Ser
          115          120          125

Gly Phe Gly Met Glu Thr Gly Gln Gly Leu Phe Asp Gly Gly Leu Phe
          130          135          140

Asp Gly Val Met Asn Pro Met Asn Leu Gly Phe Tyr Asn Gln Pro Ile
          145          150          155          160

Met Ala Ser Ala Ser Thr Ala Gly Asp Val Phe Asn Cys
          165          170

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<210> 15
 <211> 724
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G179

<400> 15
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 tcgcttgaca gagtttcatg gggtcgacaa ctctgctcag ccgacaacat catccgaaga 180
 gaagccaagg agtaagaaga agaagaaaga gagagaagcg aggtacgcgt tccagacaag 240
 aagccagggt gatatactgg atgatggata caggtggagg aagtacggcc aaaaagcagt 300
 caagaacaat ccattcccca ggagctatta taagtgcaca gaagaaggat gcagagtga 360
 gaagcaagtg cagaggcaat ggggagacga aggagtgggt gtgacgacat accaagggtg 420
 tcatacacat gccgttgata aaccctctga taatttcac cacatcttga cacaaatgca 480
 catcttccct cctttttgct tgaaggaatg attagaggaa ttggattgta atattttactt 540
 tccccaaaaac gttgggctca caccatcaga cttttacttt taaactagca gcaactcaca 600
 tatctcaaaa atactaatcc ttatctttgt ctttatggga cctttgaatc catctgcttt 660
 ggtgtcttag tctcggtgc cctgtaatcg aaagtatatt catcatcaaa ttaccaaaca 720
 taaa 724

<210> 16
 <211> 147
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G179

<400> 16
 Met Glu Asp Arg Arg Cys Asp Val Leu Phe Pro Cys Ser Ser Ser Val
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 Asp Pro Arg Leu Thr Glu Phe His Gly Val Asp Asn Ser Ala Gln Pro
 20 25 30
 Thr Thr Ser Ser Glu Glu Lys Pro Arg Ser Lys Lys Lys Lys Glu
 35 40 45
 Arg Glu Ala Arg Tyr Ala Phe Gln Thr Arg Ser Gln Val Asp Ile Leu
 50 55 60
 Asp Asp Gly Tyr Arg Trp Arg Lys Tyr Gly Gln Lys Ala Val Lys Asn
 65 70 75 80
 Asn Pro Phe Pro Arg Ser Tyr Tyr Lys Cys Thr Glu Glu Gly Cys Arg
 85 90 95
 Val Lys Lys Gln Val Gln Arg Gln Trp Gly Asp Glu Gly Val Val Val
 100 105 110
 Thr Thr Tyr Gln Gly Val His Thr His Ala Val Asp Lys Pro Ser Asp
 115 120 125

Asn Phe His His Ile Leu Thr Gln Met His Ile Phe Pro Pro Phe Cys
 130 135 140

Leu Lys Glu
 145

<210> 17
 <211> 964
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G28

<400> 17
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 acttactagg agaatcggag ccgatactca gtgagtcgac agcgagttcg gttactcaat 180
 cttgtgtaac cggtcagagc attaaaccgg tgtacggacg aaaccctagc tttagcaaac 240
 tgtatccttg cttcaccgag agctggggag atttgccgtt gaaagaaaac gattctgagg 300
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 cgtcttccga cgaagatcgt agctctttcc cgagtgttaa gatcgagact ccggagagtt 420
 tcgcggcggg ggattctggt ccggtcaaga aggagaagac gagtccgtgt tcggcggcgg 480
 tgacggcggc gaagggaaaag cattatagag gactgagaca aaggccgtgg gggaaaattg 540
 cggcggagat tagagatccg gcgaagaacg gagctagggt ttggttagga acgtttgaga 600
 cggcggagga cgcggcgttg gcttacgaca gagctgcttt caggatgcgt ggttcccgcg 660
 ctttgttgaa ttttccgttg agagttaatt caggagaacc cgacccggtt cgaatcaagt 720
 ccaagagatc ttctttttct tcttctaacg agaacggagc tccgaagaag aggagaacgg 780
 tggccgccgg tgggtggaatg gataagggat tgacggtgaa gtgcgagggt gttgaagtgg 840
 cacgtggcga tcgtttattg gttttataat tttgattttt ctttgttgga tgattatatg 900
 attcttcaaa aaagaagaac gttaataaaa aaattcgttt attattaaaa aaaaaaaaaa 960
 aaaa 964

<210> 18
 <211> 268
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G28

<400> 18
 Met Ser Met Thr Ala Asp Ser Gln Ser Asp Tyr Ala Phe Leu Glu Ser
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 Ile Arg Arg His Leu Leu Gly Glu Ser Glu Pro Ile Leu Ser Glu Ser
 20 25 30
 Thr Ala Ser Ser Val Thr Gln Ser Cys Val Thr Gly Gln Ser Ile Lys
 35 40 45
 Pro Val Tyr Gly Arg Asn Pro Ser Phe Ser Lys Leu Tyr Pro Cys Phe
 50 55 60
 Thr Glu Ser Trp Gly Asp Leu Pro Leu Lys Glu Asn Asp Ser Glu Asp
 65 70 75 80

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<210> 19
<211> 822
<212> DNA
<213> Arabidopsis thaliana
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agttgatgaa	gatggctaac	actgtccgca	ctggcgga	ggggacagta	agaagaaaga		180
agaaggctgt	tcacaagacc	actacaaccg	atgacaagag	gctccagagc	actcttaaga		240
gagttggagt	caattccatt	cccgccattg	aagaagttaa	catttttaag	gatgatgtag		300
tcattcagtt	cattaaccct	aaagttcaag	cttcaattgc	tgctaacaca	tgggttgtga		360
gtggtacacc	acagacgaaa	aaattgcaag	acattcttcc	tcagattatc	agccaacttg		420
gaccagataa	cttggacaac	ctgaggaagc	tagcagagca	attccagaaa	caagctccag		480
gtgcaggtga	tgtcccagca	acaattccaag	aagagagca	tgatgatgat	tgccagatc		540
ttgtagtggg	agagactttc	gagaccctg	ctactgaaga	ggctcccaa	gctgctgctt		600
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<210> 20
<211> 165
<212> PRT
<213> Arabidopsis thaliana
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<400> 20
Met Asn Arg Glu Lys Leu Met Lys Met Ala Asn Thr Val Arg Thr Gly
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Gly Lys Gly Thr Val Arg Arg Lys Lys Lys Ala Val His Lys Thr Thr
      20          25          30

Thr Thr Asp Asp Lys Arg Leu Gln Ser Thr Leu Lys Arg Val Gly Val
      35          40          45

Asn Ser Ile Pro Ala Ile Glu Val Asn Ile Phe Lys Asp Asp Val
      50          55          60

Val Ile Gln Phe Ile Asn Pro Lys Val Gln Ala Ser Ile Ala Ala Asn
  65          70          75          80

Thr Trp Val Val Ser Gly Thr Pro Gln Thr Lys Lys Leu Gln Asp Ile
      85          90          95

Leu Pro Gln Ile Ile Ser Gln Leu Gly Pro Asp Asn Leu Asp Asn Leu
      100          105          110

Arg Lys Leu Ala Glu Gln Phe Gln Lys Gln Ala Pro Gly Ala Gly Asp
      115          120          125

Val Pro Ala Thr Ile Gln Glu Glu Asp Asp Asp Asp Val Pro Asp
      130          135          140

Leu Val Val Gly Glu Thr Phe Glu Thr Pro Ala Thr Glu Glu Ala Pro
  145          150          155          160

Lys Ala Ala Ala Ser
      165

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<220>
<223> G19

<400> 21
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<210> 22
<211> 248
<212> PRT
<213> Arabidopsis thaliana
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			20					25					30			
Ala	Ser	Ala	Ala	Asp	Asp	Phe	Trp	Gly	Phe	Tyr	Ser	Thr	Ser	Lys	Leu	
		35					40					45				
His	Pro	Thr	Asn	Gln	Val	Asn	Val	Lys	Glu	Glu	Ala	Val	Lys	Lys	Glu	
	50					55					60					
Gln	Ala	Thr	Glu	Pro	Gly	Lys	Arg	Arg	Lys	Arg	Lys	Asn	Val	Tyr	Arg	
65					70					75					80	
Gly	Ile	Arg	Lys	Arg	Pro	Trp	Gly	Lys	Trp	Ala	Ala	Glu	Ile	Arg	Asp	
				85					90					95		
Pro	Arg	Lys	Gly	Val	Arg	Val	Trp	Leu	Gly	Thr	Phe	Asn	Thr	Ala	Glu	
			100					105					110			
Glu	Ala	Ala	Met	Ala	Tyr	Asp	Val	Ala	Ala	Lys	Gln	Ile	Arg	Gly	Asp	
		115					120					125				
Lys	Ala	Lys	Leu	Asn	Phe	Pro	Asp	Leu	His	His	Pro	Pro	Pro	Pro	Asn	
	130					135					140					
Tyr	Thr	Pro	Pro	Pro	Ser	Ser	Pro	Arg	Ser	Thr	Asp	Gln	Pro	Pro	Ala	
145					150					155					160	

Lys Lys Val Cys Val Val Ser Gln Ser Glu Ser Glu Leu Ser Gln Pro
 165 170 175
 Ser Phe Pro Val Glu Cys Ile Gly Phe Gly Asn Gly Asp Glu Phe Gln
 180 185 190
 Asn Leu Ser Tyr Gly Phe Glu Pro Asp Tyr Asp Leu Lys Gln Gln Ile
 195 200 205
 Ser Ser Leu Glu Ser Phe Leu Glu Leu Asp Gly Asn Thr Ala Glu Gln
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 Pro Ser Gln Leu Asp Glu Ser Val Ser Glu Val Asp Met Trp Met Leu
 225 230 235 240
 Asp Asp Val Ile Ala Ser Tyr Glu
 245

<210> 23
 <211> 914
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G503

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 aaaaaaaaaa aaaa 914

<210> 24
 <211> 268
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G503

<400> 24
 Met Glu Val Thr Ser Gln Ser Thr Leu Pro Pro Gly Phe Arg Phe His
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Pro Thr Asp Glu Glu Leu Ile Val Tyr Tyr Leu Arg Asn Gln Thr Met
 20 25 30
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 35 40 45
 Phe Asp Pro Trp Gln Leu Pro Glu Lys Thr Glu Phe Gly Glu Asn Glu
 50 55 60
 Trp Tyr Phe Phe Ser Pro Arg Glu Arg Lys Tyr Pro Asn Gly Val Arg
 65 70 75 80
 Pro Asn Arg Ala Ala Val Ser Gly Tyr Trp Lys Ala Thr Gly Thr Asp
 85 90 95
 Lys Ala Ile His Ser Gly Ser Ser Asn Val Gly Val Lys Lys Ala Leu
 100 105 110
 Val Phe Tyr Lys Gly Arg Pro Pro Lys Gly Ile Lys Thr Asp Trp Ile
 115 120 125
 Met His Glu Tyr Arg Leu His Asp Ser Arg Lys Ala Ser Thr Lys Arg
 130 135 140
 Ser Gly Ser Met Arg Leu Asp Glu Trp Val Leu Cys Arg Ile Tyr Lys
 145 150 155 160
 Lys Arg Gly Ala Ser Lys Leu Leu Asn Glu Gln Glu Gly Phe Met Asp
 165 170 175
 Glu Val Leu Met Glu Asp Glu Thr Lys Val Val Ile Asn Glu Ala Glu
 180 185 190
 Arg Arg Asn Asp Glu Glu Ile Met Met Met Thr Ser Met Lys Leu Pro
 195 200 205
 Arg Thr Cys Ser Leu Ala His Leu Leu Glu Met Asp Tyr Met Gly Pro
 210 215 220
 Val Ser His Ile Asp Asn Phe Ser Gln Phe Asp His Leu His Gln Pro
 225 230 235 240
 Asp Ser Glu Ser Ser Trp Phe Gly Asp Leu Gln Phe Asn Gln Asp Glu
 245 250 255
 Ile Leu Asn His His Arg Gln Ala Met Phe Lys Phe
 260 265

<210> 25
 <211> 1121
 <212> DNA
 <213> Arabidopsis thaliana

 <220>
 <223> G263

 <400> 25


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aaatgattta aaaaactatt tttttccgta aggaagaaaa gttattttta tgttttaaaa 1020
agggtgaagaa ggtccagaag gatcaacgca aatatataaa tggattttca tgtattatat 1080
aatttaatta gtgtattaag aaaataaaac aaaaaaaaaa a 1121

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<210> 26
<211> 284
<212> PRT
<213> Arabidopsis thaliana

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<220>
<223> G263

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<400> 26
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  1              5              10              15

Ser Lys Thr Tyr Gln Leu Val Asp Asp His Ser Thr Asp Asp Val Val
          20              25              30

Ser Trp Asn Glu Glu Gly Thr Ala Phe Val Val Trp Lys Thr Ala Glu
          35              40              45

Phe Ala Lys Asp Leu Leu Pro Gln Tyr Phe Lys His Asn Asn Phe Ser
          50              55              60

Ser Phe Ile Arg Gln Leu Asn Thr Tyr Gly Phe Arg Lys Thr Val Pro
          65              70              75              80

Asp Lys Trp Glu Phe Ala Asn Asp Tyr Phe Arg Arg Gly Gly Glu Asp
          85              90              95

Leu Leu Thr Asp Ile Arg Arg Arg Lys Ser Val Ile Ala Ser Thr Ala
          100             105             110

Gly Lys Cys Val Val Val Gly Ser Pro Ser Glu Ser Asn Ser Gly Gly
          115             120             125

Gly Asp Asp His Gly Ser Ser Ser Thr Ser Ser Pro Gly Ser Ser Lys
          130             135             140

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<400> 27						
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tcagtactca	tcctctttgg	tcgatacttc	attagatctc	actattggcg	ttactcgtat	180
gcgagttgaa	gaagatccac	cgacaagtgc	tttgggtgaa	gaattaaacc	gagttagtgc	240
tgagaacaag	aagctctcgg	agatgctaac	tttgtagtgt	gacacataca	acgtcttagag	300
gaagcaactt	atggaatatg	ttaacaagag	caacataaac	gagagggatc	aaatcagccc	360
tcccaagaaa	cgcaaatccc	cggcgagaga	ggacgcattc	agctgcgcgg	ttattggcgg	420
agtgtcggag	agtagctcaa	ggatcaaga	tgagtatttg	tgtaagaagc	agagagaaga	480
gactgtcgtg	aaggagaaag	tctcaagggt	ctattacaag	accgaagctt	ctgacactac	540
cctcgttgtg	aaagatgggt	atcaatggag	gaaatatgga	cagaaagtga	ctagagacaa	600
tccatctcca	agagcttact	tcaaatgtgc	ttgtgctcca	agctgtttctg	tcaaaaagaa	660
ggttcagaga	agtgtggagg	atcagtcctg	gttagttgca	acttatgagg	gtgaacacaa	720
ccatccaatg	ccatcgcaga	tcgattcaaa	caatggctta	aaccgccaca	tctctcatgg	780
tggttcagct	tcaacaccgc	ttgcagaaa	cagagaagt	agcttgactg	tgccggtgac	840
taccgtatag	atgattgaat	cgaagaaaag	gacgagccca	acgtcaagaa	tcgattttcc	900
ccaagttcag	aaacttttgg	tggagcaaat	ggcttcttcc	ttaaccaaag	atcctaactt	960
tacaqcagct	ttagcagcag	ctgttaccgg	aaaattgtat	caacagaatc	ataccgagaa	1020

atagtttagc ttcaaattcc gttagagttt ttagatttga atttgatcatg agtaagagaa 1080
 agagagtaga ttataatccn ttgtgatact gaaaaaaaaa aaaaaaaaaa 1130

<210> 28
 <211> 302
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G291

<400> 28
 Met Asp Gln Tyr Ser Ser Ser Leu Val Asp Thr Ser Leu Asp Leu Thr
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 Ile Gly Val Thr Arg Met Arg Val Glu Glu Asp Pro Pro Thr Ser Ala
 20 25 30
 Leu Val Glu Glu Leu Asn Arg Val Ser Ala Glu Asn Lys Lys Leu Ser
 35 40 45
 Glu Met Leu Thr Leu Met Cys Asp Asn Tyr Asn Val Leu Arg Lys Gln
 50 55 60
 Leu Met Glu Tyr Val Asn Lys Ser Asn Ile Thr Glu Arg Asp Gln Ile
 65 70 75 80
 Ser Pro Pro Lys Lys Arg Lys Ser Pro Ala Arg Glu Asp Ala Phe Ser
 85 90 95
 Cys Ala Val Ile Gly Gly Val Ser Glu Ser Ser Ser Thr Asp Gln Asp
 100 105 110
 Glu Tyr Leu Cys Lys Lys Gln Arg Glu Glu Thr Val Val Lys Glu Lys
 115 120 125
 Val Ser Arg Val Tyr Tyr Lys Thr Glu Ala Ser Asp Thr Thr Leu Val
 130 135 140
 Val Lys Asp Gly Tyr Gln Trp Arg Lys Tyr Gly Gln Lys Val Thr Arg
 145 150 155 160
 Asp Asn Pro Ser Pro Arg Ala Tyr Phe Lys Cys Ala Cys Ala Pro Ser
 165 170 175
 Cys Ser Val Lys Lys Lys Val Gln Arg Ser Val Glu Asp Gln Ser Val
 180 185 190
 Leu Val Ala Thr Tyr Glu Gly Glu His Asn His Pro Met Pro Ser Gln
 195 200 205
 Ile Asp Ser Asn Asn Gly Leu Asn Arg His Ile Ser His Gly Gly Ser
 210 215 220
 Ala Ser Thr Pro Val Ala Ala Asn Arg Arg Ser Ser Leu Thr Val Pro
 225 230 235 240

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<210> 29
<211> 748
<212> DNA
<213> Arabidopsis thaliana
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<400> 29						
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tttccggagc	tagactttgtc	agatgaatgg	atggatgatg	atcttgtgtc	tgcggtttcc	180
gggatgaatc	agtcttatgg	ttatcacagt	agtgatgttg	ctgggtgctt	attctcaggt	240
tctctagtct	gtttcagtcg	tcctgaatct	ccaagtacca	aaacttatgt	tgctgctaca	300
gccactgctt	ctgccgacaa	ccaaaacaag	aaagaaaaga	aaaaaattaa	agggagagtt	360
gcgttcaaga	cacggtccga	ggtggaagtg	cttgacgacg	ggttcaagtg	gagaaagtat	420
gggaagaaga	tggtgaagaa	cagcccacat	ccaagaaact	actacaaatg	ttcagttgat	480
ggctgtcccg	tgaagaaaag	ggttgaacga	gacagagatg	atccgagctt	tgtgataaca	540
acttacgagg	gttcccacaa	tcactcaagc	atgaactaag	actcgaacta	aggtcaagg	600
cgaccatgct	atattcagca	catcttattt	tctaaggtta	cgaacgatac	ttaaaaactgc	660
ttctagtctt	ttatatccat	tgtaaaactgg	ttgcaggttc	acaaaatttg	agaggtttat	720
gacattctaa	atctgtagta	cttatata				748

<220>
<223> G1275

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<400> 30
Met Asn Asp Ala Asp Thr Asn Leu Gly Ser Ser Phe Ser Asp Asp Thr
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His Ser Val Phe Glu Phe Pro Glu Leu Asp Leu Ser Asp Glu Trp Met
      20                25                30

Asp Asp Asp Leu Val Ser Ala Val Ser Gly Met Asn Gln Ser Tyr Gly
  35                40                45

Tyr Gln Thr Ser Asp Val Ala Gly Ala Leu Phe Ser Gly Ser Ser Ser
  50                55                60

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<210> 31
<211> 1195
<212> DNA
<213> Arabidopsis thaliana
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<400>	31						
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ggcttgttgt	taaatacggg	ccaagaaact	ggacagtgat	tagcaaatct	attcccggta	180	
gatcggggaa	atcgtgtcgt	ttacgggtgt	gcaaccagat	ttcgccgcaa	gttgagcatc	240	
ggcggtttct	ggctgaggaa	gacgagacga	tcgcacgtgc	tcacgctcag	ttcggggaata	300	
aatggcgcac	gattgtctcg	cttctcaacg	gtcgtacgga	caacgccgtg	agaatcact	360	
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ttggagattgg	gagaatcgag	tagacaaagt	gagattatta	ggaaactgtt	taaattggag	1020	
aagaagaaaa	atgctctgtt	tttttctcct	ttggattagg	cttaagaatt	ttgggttttta	1080	
aggaaaatgta	ttagggaaat	cgagtgaaaca	aagctcgaga	gctgggggacg	tagtgacgaa	1140	
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<210> 32
<211> 305
<212> PRT
<213> Arabidopsis thaliana
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<223> G242

Met Ala Asp Arg Ile Lys Gly Pro Trp Ser Pro Glu Glu Asp Glu Gln
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Leu Arg Arg Leu Val Val Lys Tyr Gly Pro Arg Asn Trp Thr Val Ile
20 25 30

Ser Lys Ser Ile Pro Gly Arg Ser Gly Lys Ser Cys Arg Leu Arg Trp
35 40 45

Cys Asn Gln Leu Ser Pro Gln Val Glu His Arg Pro Phe Ser Ala Glu
50 55 60

Glu Asp Glu Thr Ile Ala Arg Ala His Ala Gln Phe Gly Asn Lys Trp
65 70 75 80

Ala Thr Ile Ala Arg Leu Leu Asn Gly Arg Thr Asp Asn Ala Val Lys
85 90 95

Asn His Trp Asn Ser Thr Leu Lys Arg Lys Cys Gly Gly Tyr Asp His
100 105 110

Arg Gly Tyr Asp Gly Ser Glu Asp His Arg Pro Val Lys Arg Ser Val
115 120 125

Ser Ala Gly Ser Pro Pro Val Val Thr Gly Leu Tyr Met Ser Pro Gly
130 135 140

Ser	Pro	Thr	Gly	Ser	Asp	Val	Ser	Asp	Ser	Ser	Thr	Ile	Pro	Ile	Leu
145					150					155					160

Pro Ser Val Glu Leu Phe Lys Pro Val Pro Arg Pro Gly Ala Val Val
165 170 175

Leu Pro Leu Pro Ile Glu Thr Ser Ser Phe Ser Asp Asp Pro Pro Thr
180 185 190

Ser Leu Ser Leu Ser Leu Pro Gly Ala Asp Val Ser Glu Glu Ser Asn
195 200 205

Arg Ser His Glu Ser Thr Asn Ile Asn Asn Thr Thr Ser Ser Arg His
210 215 220

Asn His Asn Asn Thr Val Ser Phe Met Pro Phe Ser Gly Gly Phe Arg
225 230 235 240

Gly Ala Ile Glu Glu Met Gly Lys Ser Phe Pro Gly Asn Gly Gly Glu
245 250 255

Phe Met Ala Val Val Gln Glu Met Ile Lys Ala Glu Val Arg Ser Tyr
260 265 270

Met Thr Glu Met Gln Arg Asn Asn Gly Gly Gly Phe Val Gly Gly Phe
275 280 285

Glu
305

<220>
<223> G1006

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ctcaaagatg	ccttccattt	tgacacgtca	tcatacggact	tgagctgtct	ttttgatttt	300	
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aaaaaaaaaa	aaa					913	

<220>
<223> G1006

<400> 34																
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Ile	Thr	Arg	His	Leu	Leu	Gly	Gly	Gly	Gly	Glu	Asn	Glu	Leu	Arg	Leu	
			20					25					30			
Asn	Glu	Ser	Thr	Pro	Ser	Ser	Cys	Phe	Thr	Glu	Ser	Trp	Gly	Gly	Leu	
		35					40					45				
Pro	Leu	Lys	Glu	Asn	Asp	Ser	Glu	Asp	Met	Leu	Val	Tyr	Gly	Leu	Leu	
	50					55					60					
Lys	Asp	Ala	Phe	His	Phe	Asp	Thr	Ser	Ser	Ser	Asp	Leu	Ser	Cys	Leu	
65					70					75					80	

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<210> 35
<211> 725
<212> DNA
<213> Arabidopsis thaliana
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gccattttcca	accaacgggtc	aaaacccgta	cctcctctac	ggattccaaa	gcctacaaa	180
caatccacaa	tccatgagcc	taagcagcaa	caactcaaca	tcagatgaag	cagaagagca	240
gcagacgaac	aacaatataa	tcaacgagcg	gaagcagaga	aggatgattt	caaaccgaga	300
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gatgtgggtta	aggatcgaga	atcatcagtt	gcttgataag	cttaacaatc	tctctgagtc	420
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cattgaataa	agcattttttc	cccgattcat	atztatgaaa	attttcttca	agagtatggt	600
tctttgtatg	tatatgtgga	gatgtatttc	agggttttga	taatatgacc	ctttacgacg	660
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tataa						725

<210> 36
 <211> 173
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G1049

<400> 36
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 20 25 30
 Phe Pro Thr Asn Gly Gln Asn Pro Tyr Leu Leu Tyr Gly Phe Gln Ser
 35 40 45
 Pro Thr Asn Asn Pro Gln Ser Met Ser Leu Ser Ser Asn Asn Ser Thr
 50 55 60
 Ser Asp Glu Ala Glu Glu Gln Gln Thr Asn Asn Asn Ile Ile Asn Glu
 65 70 75 80
 Arg Lys Gln Arg Arg Met Ile Ser Asn Arg Glu Ser Ala Arg Arg Ser
 85 90 95
 Arg Met Arg Lys Gln Arg His Leu Asp Glu Leu Trp Ser Gln Val Met
 100 105 110
 Trp Leu Arg Ile Glu Asn His Gln Leu Leu Asp Lys Leu Asn Asn Leu
 115 120 125
 Ser Glu Ser His Asp Lys Val Leu Gln Glu Asn Ala Gln Leu Lys Glu
 130 135 140
 Glu Thr Phe Glu Leu Lys Gln Val Ile Ser Asp Met Gln Ile Gln Ser
 145 150 155 160
 Pro Phe Ser Cys Phe Arg Asp Asp Ile Ile Pro Ile Glu
 165 170

<210> 37
 <211> 1409
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G502

<400> 37
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 cgcgtccgaa ttgattagga taggatcagg atcatcctca acaacctcct cctaattcct 180
 cctccattca tagtaacaat aatattaaga aagagggtaa actatgtcag aattattaca 240
 gttgcctcca ggtttccgat ttcaccctac cgatgaagag cttgtcatgc actatctctg 300
 ccgcaaattg gcctctcagt ccacgcgcgt tccgatcatc gctgagatcg atctctacaa 360

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gaaagctctt gttttctacg ccggcaaagc tccaaagggg gagaaaacca attggatcat 600
gcacgagtac cgtctcgccg acgttgaccg gtccgttcgc aagaagaaga atagtctcag 660
gctggatgat tgggttctct gccggattta caacaaaaaa ggagctaccg agaggcgggg 720
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gaagccttac tagaagggaa ttcctttcct gccgccgaaa cgcaacgcaa aacgaccctc 1140
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atttaatat ttctgtctaa aaaaaaaaaa 1409

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<210> 38
 <211> 319
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G502

<400> 38
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 Asp Glu Glu Leu Val Met His Tyr Leu Cys Arg Lys Cys Ala Ser Gln
 20 25 30
 Ser Ile Ala Val Pro Ile Ile Ala Glu Ile Asp Leu Tyr Lys Tyr Asp
 35 40 45
 Pro Trp Glu Leu Pro Gly Leu Ala Leu Tyr Gly Glu Lys Glu Trp Tyr
 50 55 60
 Phe Phe Ser Pro Arg Asp Arg Lys Tyr Pro Asn Gly Ser Arg Pro Asn
 65 70 75 80
 Arg Ser Ala Gly Ser Gly Tyr Trp Lys Ala Thr Gly Ala Asp Lys Pro
 85 90 95
 Ile Gly Leu Pro Lys Pro Val Gly Ile Lys Lys Ala Leu Val Phe Tyr
 100 105 110
 Ala Gly Lys Ala Pro Lys Gly Glu Lys Thr Asn Trp Ile Met His Glu
 115 120 125
 Tyr Arg Leu Ala Asp Val Asp Arg Ser Val Arg Lys Lys Lys Asn Ser
 130 135 140
 Leu Arg Leu Asp Asp Trp Val Leu Cys Arg Ile Tyr Asn Lys Lys Gly
 145 150 155 160

G502: Arabidopsis thaliana

Ala Thr Glu Arg Arg Gly Pro Pro Pro Pro Val Val Tyr Gly Asp Glu
165 170 175

Ile Met Glu Glu Lys Pro Lys Val Thr Glu Met Val Met Pro Pro Pro
180 185 190

Pro Gln Gln Thr Ser Glu Phe Ala Tyr Phe Asp Thr Ser Asp Ser Val
195 200 205

Pro Lys Leu His Thr Thr Asp Ser Ser Cys Ser Glu Gln Val Val Ser
210 215 220

Pro Glu Phe Thr Ser Glu Val Gln Ser Glu Pro Lys Trp Lys Asp Trp
225 230 235 240

Ser Ala Val Ser Asn Asp Asn Asn Asn Thr Leu Asp Phe Gly Phe Asn
245 250 255

Tyr Ile Asp Ala Thr Val Asp Asn Ala Phe Gly Gly Gly Gly Ser Ser
260 265 270

Asn Gln Met Phe Pro Leu Gln Asp Met Phe Met Tyr Met Gln Lys Pro
275 280 285

Tyr Lys Gly Ile Pro Phe Leu Pro Pro Lys Arg Asn Ala Lys Arg Pro
290 295 300

Ser Phe Leu Arg Leu Trp Gln His Glu Thr Val Leu Tyr Gly Gln
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<210> 39

<211> 1347

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G239

<400> 39

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tgtagattaa gatggcttaa ttacttacgt ccagatgtta gaagaggcaa catcactctc 240
gaagaacaat ttatgatcct caaactccat tctctttggg gcaatagggtg gtcgaagatt 300
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tgtgaacaag tggagtcaat gatcaccgac ccaagtcaac cagttaacga accgagtccg 540
gtcgagccgg gtttcgttca attcagccag aatcatcatc agcaattcgt accggctacg 600
gaattgtcag caacgtcttc gaattctccg gctgagacgt tttcggacgt tccgaggtggg 660
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aatttttagat acggccttgg tatacgaatc tttgacttaa ttattatctt ttcttttctt 960
ctcttgtttt aaacccttga ttaaattaag atttgatcat cagacgagga tatttgtgat 1020

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aaaattaaaa aacgacaaaa cgaaaatatg actaaattta tttttttgtc agttaaccac 1260
tgattatagg ttgaaattgt cacaacacat gatttatctt gatagaaatt tagtagtcca 1320
gaatgctgca tggttgatcc taagaaa                                     1347

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<210> 40
 <211> 273
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G239

<400> 40
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 Asp Ser Asp Val Arg Lys Gly Pro Trp Thr Glu Glu Glu Asp Ala Ile
 20 25 30
 Leu Val Asn Phe Val Ser Ile His Gly Asp Ala Arg Trp Asn His Ile
 35 40 45
 Ala Arg Ser Ser Gly Leu Lys Arg Thr Gly Lys Ser Cys Arg Leu Arg
 50 55 60
 Trp Leu Asn Tyr Leu Arg Pro Asp Val Arg Arg Gly Asn Ile Thr Leu
 65 70 75 80
 Glu Glu Gln Phe Met Ile Leu Lys Leu His Ser Leu Trp Gly Asn Arg
 85 90 95
 Trp Ser Lys Ile Ala Gln Tyr Leu Pro Gly Arg Thr Asp Asn Glu Ile
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 Cys Asp Val Asn Ser Asn Leu Phe Lys Glu Thr Met Arg Asn Val Trp
 130 135 140
 Met Pro Arg Leu Val Glu Arg Ile Asn Ala Gln Ser Leu Pro Thr Thr
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 Cys Glu Gln Val Glu Ser Met Ile Thr Asp Pro Ser Gln Pro Val Asn
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 Glu Pro Ser Pro Val Glu Pro Gly Phe Val Gln Phe Ser Gln Asn His
 180 185 190
 His Gln Gln Phe Val Pro Ala Thr Glu Leu Ser Ala Thr Ser Ser Asn
 195 200 205
 Ser Pro Ala Glu Thr Phe Ser Asp Val Arg Gly Gly Val Val Asn Gly
 210 215 220

Ser Gly Tyr Asp Pro Ser Gly Gln Thr Gly Phe Gly Glu Phe Asn Asp
225 230 235 240

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Arg Arg Leu Ala Gln Asn Arg Glu Ala Ala Arg Lys Ser Arg Leu Arg
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Lys Lys Ala Tyr Val Gln Gln Leu Glu Asn Ser Arg Leu Lys Leu Thr
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Gln Leu Glu Gln Glu Leu Gln Arg Ala Arg Gln Gln Gly Val Phe Ile
85 90 95

Ser Ser Ser Gly Asp Gln Ala His Ser Thr Ala Gly Asp Gly Ala Met
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Ala Phe Asp Val Glu Tyr Arg Arg Trp Gln Glu Asp Lys Asn Arg Gln
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Arg Ile Lys Gly Asn Ala Ala Lys Ser Asp Val Phe His Leu Leu Ser
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180 185 190

Phe Arg Ser Ser Glu Leu Leu Lys Leu Ile Ala Cys Gln Leu Glu Pro
195 200 205

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210 215 220

Gln Gln Ala Glu Asp Ala Leu Ser Gln Gly Met Asp Asn Leu Gln Gln
225 230 235 240

Ser Leu Ala Asp Thr Leu Ser Ser Gly Thr Leu Gly Ser Ser Ser Ser
245 250 255

Gly Asn Val Ala Ser Tyr Met Gly Gln Met Ala Met Ala Met Gly Lys
260 265 270

Leu Gly Thr Leu Glu Gly Phe Ile Arg Gln Ala Asp Asn Leu Arg Leu
275 280 285

Gln Thr Tyr Gln Gln Met Val Arg Leu Leu Thr Thr Arg Gln Ser Ala
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Tyr Lys Cys Thr Val Cys Gly Lys Ser Phe Ser Ser Tyr Gln Ala Leu
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 115 120 125
 Ser Gly Asn Gln Glu Leu Ser Asn Asn Ser His Ser Asn Ser Gly Ser
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 Gly Lys Ile His Thr Cys Ser Ile Cys Phe Lys Ser Phe Ala Ser Gly
 165 170 175
 Gln Ala Leu Gly Gly His Lys Arg Cys His Tyr Asp Gly Gly Asn Asn
 180 185 190
 Gly Asn Gly Asn Gly Ser Ser Ser Asn Ser Val Glu Leu Val Ala Gly
 195 200 205
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<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G553

<400> 50

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Arg Asp Met Gly Met Tyr Glu Pro Phe Gln Gln Leu Ser Gly Trp Glu
      20              25              30

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Ser Pro Phe Lys Ser Asp Ile Asn Asn Ile Thr Ser Asn Gln Asn Asn
      35              40              45

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Asn Gln Ser Ser Ser Thr Thr Leu Glu Val Asp Ala Arg Pro Glu Ala
      50              55              60

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Asp Asp Asn Asn Arg Val Asn Tyr Thr Ser Val Tyr Asn Asn Ser Leu
      65              70              75              80

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Glu Ala Glu Pro Ser Ser Asn Asn Asp Gln Asp Glu Asp Arg Ile Asn
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<210> 51
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<212> DNA
<213> Arabidopsis thaliana

<220>
<223> G1221

<400> 51

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<213> Arabidopsis thaliana

<220>
<223> G1221

<400> 52

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Lys Ser Ile Pro Pro Trp Lys Glu Gln Ile Thr Phe Arg Gly Ile Val
35 40 45

Ala Ser Leu Ile Ile Gly Ile Ile Tyr Ser Val Ile Val Met Lys Leu
50 55 60

Asn Leu Thr Thr Gly Leu Val Pro Asn Leu Asn Val Ser Ala Ala Leu
65 70 75 80

Leu Ala Phe Val Phe Leu Arg Ser Trp Thr Lys Leu Leu Thr Lys Ala
85 90 95

Gly Ile Val Thr Lys Pro Phe Thr Lys Gln Glu Asn Thr Val Val Gln
100 105 110

Thr Cys Ala Val Ala Cys Tyr Ser Ile Ala Val Gly Gly Gly Phe Gly
115 120 125

Ser Tyr Leu Leu Gly Leu Asn Arg Ile Thr Tyr Glu Gln Ser Gly Gly
130 135 140

Thr His Thr Asp Gly Asn Tyr Pro Glu Gly Thr Lys Glu Pro Gly Ile
145 150 155 160

Gly Trp Met Thr Ala Phe Leu Phe Phe Thr Cys Phe Val Gly Leu Leu
165 170 175

Ala Leu Val Pro Leu Arg Lys Ile Met Ile Ile Asp Tyr Lys Leu Thr
180 185 190

Tyr Pro Ser Gly Thr Ala Thr Ala Val Leu Ile Asn Gly Phe His Thr
195 200 205

Pro Lys Gly Asn Lys Met Ala Lys Lys Gln Val Phe Gly Phe Val Lys
210 215 220

Tyr Phe Ser Phe Ser Phe Ile Trp Ala Phe Phe Gln Trp Phe Phe Ser
225 230 235 240

Gly Gly Thr Glu Cys Gly Phe Ile Gln Phe Pro Thr Phe Gly Leu Glu
245 250 255

Ala Leu Lys Asn Thr Phe Tyr Phe Asp Phe Ser Met Thr Tyr Val Gly
260 265 270

Ala Gly Met Ile Cys Pro His Ile Val Asn Ile Ser Leu Leu Phe Gly
275 280 285

Ala Val Leu Ser Trp Gly Ile Met Trp Pro Leu Ile Lys Gly Leu Lys
290 295 300

Gly 305	Asp	Trp	Phe	Pro	Ser 310	Thr	Leu	Pro	Glu	Asn 315	Ser	Met	Lys	Ser	Leu 320
Asn	Gly	Tyr	Lys	Val 325	Phe	Ile	Ser	Ile	Ser 330	Leu	Ile	Leu	Gly	Asp 335	Gly
Leu	Tyr	Gln	Phe 340	Ile	Lys	Ile	Leu	Phe 345	Lys	Thr	Gly	Ile	Asn 350	Met	Tyr
Val	Lys	Leu 355	Asn	Asn	Arg	Asn	Ser 360	Gly	Lys	Ser	Asn	Ser 365	Glu	Lys	Asp
Lys	Gln 370	Ser	Ile	Ala	Asp	Leu 375	Lys	Arg	Asp	Glu	Ile 380	Phe	Val	Arg	Asp
Ser 385	Ile	Pro	Leu	Trp	Val 390	Ala	Ala	Val	Gly	Asn 395	Ala	Ala	Phe	Ser	Val 400
Val	Ser	Ile	Ile 405	Ala	Ile	Pro	Ile	Met	Phe 410	Pro	Glu	Leu	Lys	Trp 415	Tyr
Phe	Ile	Val 420	Val	Ala	Tyr	Met	Leu	Ala 425	Pro	Ser	Leu	Gly	Phe 430	Ser	Asn
Ala	Tyr 435	Gly	Ala	Gly	Leu	Thr	Asp 440	Met	Asn	Met	Ala	Tyr 445	Asn	Tyr	Gly
Lys	Val 450	Ala	Leu	Phe	Ile	Leu 455	Ala	Ala	Met	Ala	Gly 460	Lys	Gln	Asn	Gly
Val 465	Val	Ala	Gly	Leu 470	Val	Gly	Cys	Gly	Leu	Ile 475	Lys	Ser	Ile	Val	Ser 480
Ile	Ser	Ser	Asp 485	Leu	Met	His	Asp	Phe	Lys 490	Thr	Gly	His	Leu	Thr 495	Leu
Thr	Ser	Pro	Arg 500	Ser	Met	Leu	Val	Ser 505	Gln	Ala	Ile	Gly	Thr 510	Ala	Ile
Gly	Cys 515	Val	Val	Ala	Pro	Leu	Thr 520	Phe	Phe	Leu	Phe	Tyr 525	Lys	Ala	Phe
Asp	Val 530	Gly	Asn	Gln	Glu	Gly 535	Glu	Tyr	Lys	Ala	Pro 540	Tyr	Ala	Leu	Val
Tyr 545	Arg	Asn	Met	Ala 550	Ile	Leu	Gly	Val	Glu	Gly 555	Phe	Ser	Ala	Leu	Pro 560
Gln	His	Cys	Leu 565	Gln	Leu	Cys	Tyr	Gly	Phe 570	Phe	Ala	Phe	Ala	Val 575	Ala
Ala	Asn	Leu 580	Val	Arg	Asp	Arg	Leu	Pro 585	Asp	Lys	Ile	Gly	Asn 590	Trp	Val
Pro	Leu 595	Pro	Met	Ala	Met	Ala 600	Val	Pro	Phe	Leu	Val 605	Gly	Gly	Tyr	Phe

Ala Ile Asp Met Cys Val Gly Ser Leu Ile Val Phe Ala Trp Asn Met
610 615 620

Arg Asp Arg Val Lys Ala Gly Leu Met Val Pro Ala Val Ala Ser Gly
625 630 635 640

Leu Ile Cys Gly Asp Gly Leu Trp Ile Leu Pro Ser Ser Val Leu Ala
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Tyr Ser Ser
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<212> DNA

<213> Arabidopsis thaliana

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<223> G580

<400> 53

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<211> 234

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G580

<400> 54

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 35 40 45
 Leu Gly Ser Leu His Tyr His Arg Gln Leu Asn Ile Gly His Glu Pro
 50 55 60
 Met Leu Lys Asn Gln Asn Pro Asn Asn Ser Ile Phe Gln Asp Phe Leu
 65 70 75 80
 Asn Met Pro Leu Asn Gln Pro Pro Pro Pro Pro Pro Pro Pro Ser Ser
 85 90 95
 Ser Thr Ile Val Thr Ala Leu Tyr Gly Ser Leu Pro Leu Pro Pro Pro
 100 105 110
 Ala Thr Val Leu Ser Leu Asn Ser Gly Val Gly Phe Glu Phe Leu Asp
 115 120 125
 Thr Thr Glu Asn Leu Leu Ala Ser Asn Pro Arg Ser Phe Glu Glu Ser
 130 135 140
 Ala Lys Phe Gly Cys Leu Gly Lys Lys Arg Gly Gln Asp Ser Asp Asp
 145 150 155 160
 Thr Arg Gly Asp Arg Arg Tyr Lys Arg Met Ile Lys Asn Arg Glu Ser
 165 170 175
 Ala Ala Arg Ser Arg Ala Arg Lys Gln Ala Tyr Thr Asn Glu Leu Glu
 180 185 190
 Leu Glu Ile Ala His Leu Gln Thr Glu Asn Ala Arg Leu Lys Ile Gln
 195 200 205
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 210 215 220
 Thr Leu Gln Arg Ser Ser Thr Ala Pro Phe
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<210> 55

<211> 1575

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G270

<400> 55

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<210> 56
<211> 435
<212> PRT
<213> Arabidopsis thaliana

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<220>
<223> G270

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                20                25                30

Thr Arg Leu Arg Ser Leu Ser Tyr Ser Ser Gln Thr Ser Ile Leu Pro
                35                40                45

Asp Ala Gly Asp Asp Phe Ile Val Gly Asp Cys Leu Val Tyr Glu Asp
 50                55                60

Gly Val Phe Glu Asp Pro Tyr Leu Asp Lys Glu Val Thr Gln Val Ala
 65                70                75                80

Lys Gln Glu Arg Lys Lys Asn Arg Arg Gly Gly Ala Lys Arg Leu Asp
                85                90                95

Glu Ser Glu Ile Glu Pro Glu Asn Leu Val Pro Glu Glu Trp Arg Asp
                100                105                110

Ile Gln Ala Glu Val Asn Leu Thr Lys Lys Asp Lys Arg Lys Ile Ala
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Gln Glu Met Glu Phe Gly Val Arg Val Glu Lys Lys Arg Gln Gly Leu
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 <223> G201

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<210> 58
 <211> 336
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G201

<400> 58
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 35 40 45
 Cys Gly Lys Ser Cys Arg Leu Arg Trp Ala Asn Tyr Leu Lys Pro Asp
 50 55 60
 Ile Lys Arg Gly Glu Phe Ser Tyr Glu Glu Glu Gln Ile Ile Ile Met
 65 70 75 80

Leu His Ala Ser Arg Gly Asn Lys Trp Ser Val Ile Ala Arg His Leu
 85 90 95
 Pro Lys Arg Thr Asp Asn Glu Ile Lys Asn Tyr Trp Asn Thr His Leu
 100 105 110
 Lys Lys Leu Leu Ile Asp Lys Gly Ile Asp Pro Val Thr His Lys Pro
 115 120 125
 Leu Ala Tyr Asp Ser Asn Pro Asp Glu Gln Ser Gln Ser Gly Ser Ile
 130 135 140
 Ser Pro Lys Ser Leu Pro Pro Ser Ser Ser Lys Asn Val Pro Glu Ile
 145 150 155 160
 Thr Ser Ser Asp Glu Thr Pro Lys Tyr Asp Ala Ser Leu Ser Ser Lys
 165 170 175
 Lys Arg Cys Phe Lys Arg Ser Ser Ser Thr Ser Lys Leu Leu Asn Lys
 180 185 190
 Val Ala Ala Arg Ala Ser Ser Met Gly Thr Ile Leu Gly Ala Ser Ile
 195 200 205
 Glu Gly Thr Leu Ile Ser Ser Thr Pro Leu Ser Ser Cys Leu Asn Asp
 210 215 220
 Asp Phe Ser Glu Thr Ser Gln Phe Gln Met Glu Glu Phe Asp Pro Phe
 225 230 235 240
 Tyr Gln Ser Ser Glu His Ile Ile Asp His Met Lys Glu Asp Ile Ser
 245 250 255
 Ile Asn Asn Ser Glu Tyr Asn Phe Ser Gln Phe Leu Glu Gln Phe Ser
 260 265 270
 Asn Asn Glu Gly Glu Glu Ala Asp Asn Thr Gly Gly Gly Tyr Asn Gln
 275 280 285
 Asp Leu Leu Met Ser Asp Val Ser Ser Thr Ser Val Asp Glu Asp Glu
 290 295 300
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<210> 59

<211> 1651

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G1417

<400> 59


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tagccctcc gccacgtcat catcatcctt ctaccataac ttcccataca cctccacaat 1080
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tcgaccgcta caaccgccac cgcagtttct aagccagtat ggtcccgcg cgtttttacc 1200
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gagtagtaac ggagattcgc cacagcttcc tcagtcttgc accactttct ctacaaacta 1500
attttactac cattattata tgttatctta ttatatatta cacacacata ttatacatta 1560
tgcgtatctt aagttttttt ttgggggcca ttatatatga atgatatgga gatcactgag 1620
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<210> 60

<211> 489

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G1417

<400> 60

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Met Glu Glu His Ile Gln Asp Arg Arg Glu Ile Ala Phe Leu His Ser
  1             5             10            15

Gly Glu Phe Leu His Gly Asp Ser Asp Ser Lys Asp His Gln Pro Asn
      20             25             30

Glu Ser Pro Val Glu Arg His His Glu Ser Ser Ile Lys Glu Val Asp
      35             40             45

Phe Phe Ala Ala Lys Ser Gln Pro Phe Asp Leu Gly His Val Arg Thr
      50             55             60

Thr Thr Ile Val Gly Ser Ser Gly Phe Asn Asp Gly Leu Gly Leu Val
      65             70             75             80

Asn Ser Cys His Gly Thr Ser Ser Asn Asp Gly Asp Asp Lys Thr Lys
      85             90             95

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Asn Asn Gln Gln Leu Leu Ile Pro Asn Leu Phe Gly Pro Gln Ala Pro
 405 410 415
 Pro Arg Glu Met Val Asp Ser Val Arg Ala Ala Ile Ala Met Asp Pro
 420 425 430
 Asn Phe Thr Ala Ala Leu Ala Ala Ala Ile Ser Asn Ile Ile Gly Gly
 435 440 445
 Gly Asn Asn Asp Asn Asn Asn Asn Thr Asp Ile Asn Asp Asn Lys Val
 450 455 460
 Asp Ala Lys Ser Gly Gly Ser Ser Asn Gly Asp Ser Pro Gln Leu Pro
 465 470 475 480
 Gln Ser Cys Thr Thr Phe Ser Thr Asn
 485

<210> 61
 <211> 1046
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G233

<400> 61
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 ggcaatagat ggtcagcgat tgcaagcaaaa ctgcctggaa gaaccgataa cgagatcaag 360
 aacgtatggc aactcactt gaagaagaga ctogaagatt atcaaccagc taaacctaa 420
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 tcgacatcgc cttcgacaag tgaggtttct tcgatgacac tcataagcca cgacggctat 600
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 gtttctttcg aaacttttgg tgcggatatc gatgaaagct tctggaaaga gacactgtat 720
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 caacaagagt ttcaaaactt gggctccgct aataatgaga tgatttttga cagtgaatg 840
 gaacttctgg ttcgatgtat tggctagaac cggcggggaa caagatctct tagccgggct 900
 ctagttaaca tgtttgagga gtaaaagtga atgggtgcaa ttagttaagg ctaagaaatt 960
 caaaagcttt tgtttaccga gaaaaaaaca cactctaact cttgatgtga ttagttagt 1020
 gtattaatta gaggtgcgt tttcaa 1046

<210> 62
 <211> 273
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G233

Met Gly Arg Ala Pro Cys Cys Glu Lys Met Gly Leu Lys Arg Gly Pro
1 5 10 15

Trp Thr Pro Glu Glu Asp Gln Ile Leu Val Ser Phe Ile Leu Asn His
20 25 30

Gly His Ser Asn Trp Arg Ala Leu Pro Lys Gln Ala Gly Leu Leu Arg
35 40 45

Cys Gly Lys Ser Cys Arg Leu Arg Trp Met Asn Tyr Leu Lys Pro Asp
50 55 60

Ile Lys Arg Gly Asn Phe Thr Lys Glu Glu Glu Asp Ala Ile Ile Ser
65 70 75 80

Leu His Gln Ile Leu Gly Asn Arg Trp Ser Ala Ile Ala Ala Lys Leu
85 90 95

Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Val Trp His Thr His Leu
100 105 110

Lys Lys Arg Leu Glu Asp Tyr Gln Pro Ala Lys Pro Lys Thr Ser Asn
115 120 125

Lys Lys Lys Gly Thr Lys Pro Lys Ser Glu Ser Val Ile Thr Ser Ser
130 135 140

Asn Ser Thr Arg Ser Glu Ser Glu Leu Ala Asp Ser Ser Asn Pro Ser
145 150 155 160

Gly Glu Ser Leu Phe Ser Thr Ser Pro Ser Thr Ser Glu Val Ser Ser
165 170 175

Met Thr Leu Ile Ser His Asp Gly Tyr Ser Asn Glu Ile Asn Met Asp
180 185 190

Asn Lys Pro Gly Asp Ile Ser Thr Ile Asp Gln Glu Cys Val Ser Phe
195 200 205

Glu Thr Phe Gly Ala Asp Ile Asp Glu Ser Phe Trp Lys Glu Thr Leu
210 215 220

Tyr Ser Gln Asp Glu His Asn Tyr Val Ser Asn Asp Leu Glu Val Ala
225 230 235 240

Gly Leu Val Glu Ile Gln Gln Glu Phe Gln Asn Leu Gly Ser Ala Asn
245 250 255

Asn Glu Met Ile Phe Asp Ser Glu Met Glu Leu Leu Val Arg Cys Ile
260 265 270

Gly

<210> 63

<211> 1296

<212> DNA

<213> *Arabidopsis thaliana*

<220>

<223> G920

<400> 63

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cgaatagtaa caaacacgaaa tccataaaga gaaaagttgt cgaccaactt gtcgaaggct 180
atgaattcgc tactcagctt cagcttctcc tttctcatca acactctaac cagtaccaca 240
tcgatgagac ccgtcttggt tccgggctcg gttcagtttc cgggtggcca gatcccgttg 300
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agagattagg ggttggttaag ggtaaaagag gatgctacac tagaaagacg agatcacata 540
caaggatcgt ggaagctaaa agttctgaag acagatatgc ttggaggaaa tatggacaaa 600
aggagattct taataccaca ttcccaagaa gttactttag atgcacacac aagccaacgc 660
aaggatgcaa agcaacaaag caagttcaga aacaggatca agattctgag atgttccaaa 720
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accatgtcaa tgctatgggt caagagcaag agaacaacac cagcagtggt acagcaatag 900
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caagtgctat gtgaacatcc aaatctggaa tgatgaatca gcactagggtc ttctctttga 1200
gtatgtctag tttaatgtaa ttttttgggt gtatgtttga taaaaacacc atatatactt 1260
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<210> 64

<211> 346

<212> PRT

<213> *Arabidopsis thaliana*

<220>

<223> G920

<400> 64

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Met Asp Ser Asn Ser Asn Asn Thr Lys Ser Ile Lys Arg Lys Val Val
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Asp Gln Leu Val Glu Gly Tyr Glu Phe Ala Thr Gln Leu Gln Leu Leu
      20              25              30

Leu Ser His Gln His Ser Asn Gln Tyr His Ile Asp Glu Thr Arg Leu
      35              40              45

Val Ser Gly Ser Gly Ser Val Ser Gly Gly Pro Asp Pro Val Asp Glu
      50              55              60

Leu Met Ser Lys Ile Leu Gly Ser Phe His Lys Thr Ile Ser Val Leu
      65              70              75              80

Asp Ser Phe Asp Pro Val Ala Val Ser Val Pro Ile Ala Val Glu Gly
      85              90              95

```

Ser Trp Asn Ala Ser Cys Gly Asp Asp Ser Ala Thr Pro Val Ser Cys
 100 105 110
 Asn Gly Gly Asp Ser Gly Glu Ser Lys Lys Lys Arg Leu Gly Val Gly
 115 120 125
 Lys Gly Lys Arg Gly Cys Tyr Thr Arg Lys Thr Arg Ser His Thr Arg
 130 135 140
 Ile Val Glu Ala Lys Ser Ser Glu Asp Arg Tyr Ala Trp Arg Lys Tyr
 145 150 155 160
 Gly Gln Lys Glu Ile Leu Asn Thr Thr Phe Pro Arg Ser Tyr Phe Arg
 165 170 175
 Cys Thr His Lys Pro Thr Gln Gly Cys Lys Ala Thr Lys Gln Val Gln
 180 185 190
 Lys Gln Asp Gln Asp Ser Glu Met Phe Gln Ile Thr Tyr Ile Gly Tyr
 195 200 205
 His Thr Cys Thr Ala Asn Asp Gln Thr His Ala Lys Thr Glu Pro Phe
 210 215 220
 Asp Gln Glu Ile Ile Met Asp Ser Glu Lys Thr Leu Ala Ala Ser Thr
 225 230 235 240
 Ala Gln Asn His Val Asn Ala Met Val Gln Glu Gln Glu Asn Asn Thr
 245 250 255
 Ser Ser Val Thr Ala Ile Asp Ala Gly Met Val Lys Glu Glu Gln Asn
 260 265 270
 Asn Asn Gly Asp Gln Ser Lys Asp Tyr Tyr Glu Gly Ser Ser Thr Gly
 275 280 285
 Glu Asp Leu Ser Leu Val Trp Gln Glu Thr Met Met Phe Asp Asp His
 290 295 300
 Gln Asn His Tyr Tyr Cys Gly Glu Thr Ser Thr Thr Ser His Gln Phe
 305 310 315 320
 Gly Phe Ile Asp Asn Asp Asp Gln Phe Ser Ser Phe Phe Asp Ser Tyr
 325 330 335
 Cys Ala Asp Tyr Glu Arg Thr Ser Ala Met
 340 345

<210> 65
 <211> 1281
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G867

<400> 65

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<210> 66
<211> 344
<212> PRT
<213> Arabidopsis thaliana
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<400> 66
Met Glu Ser Ser Ser Val Asp Glu Ser Thr Thr Ser Thr Gly Ser Ile
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Cys Glu Thr Pro Ala Ile Thr Pro Ala Lys Lys Ser Ser Val Gly Asn
      20              25              30
Leu Tyr Arg Met Gly Ser Gly Ser Ser Val Val Leu Asp Ser Glu Asn
      35              40              45
Gly Val Glu Ala Glu Ser Arg Lys Leu Pro Ser Ser Lys Tyr Lys Gly
  50              55              60
Val Val Pro Gln Pro Asn Gly Arg Trp Gly Ala Gln Ile Tyr Glu Lys
  65              70              75              80
His Gln Arg Val Trp Leu Gly Thr Phe Asn Glu Glu Asp Glu Ala Ala
      85              90              95
Arg Ala Tyr Asp Val Ala Val His Arg Phe Arg Arg Arg Asp Ala Val
      100              105              110
Thr Asn Phe Lys Asp Val Lys Met Asp Glu Asp Glu Val Asp Phe Leu
      115              120              125

```

Asn Ser His Ser Lys Ser Glu Ile Val Asp Met Leu Arg Lys His Thr
 130 135 140
 Tyr Asn Glu Glu Leu Glu Gln Ser Lys Arg Arg Arg Asn Gly Asn Gly
 145 150 155 160
 Asn Met Thr Arg Thr Leu Leu Thr Ser Gly Leu Ser Asn Asp Gly Val
 165 170 175
 Ser Thr Thr Gly Phe Arg Ser Ala Glu Ala Leu Phe Glu Lys Ala Val
 180 185 190
 Thr Pro Ser Asp Val Gly Lys Leu Asn Arg Leu Val Ile Pro Lys His
 195 200 205
 His Ala Glu Lys His Phe Pro Leu Pro Ser Ser Asn Val Ser Val Lys
 210 215 220
 Gly Val Leu Leu Asn Phe Glu Asp Val Asn Gly Lys Val Trp Arg Phe
 225 230 235 240
 Arg Tyr Ser Tyr Trp Asn Ser Ser Gln Ser Tyr Val Leu Thr Lys Gly
 245 250 255
 Trp Ser Arg Phe Val Lys Glu Lys Asn Leu Arg Ala Gly Asp Val Val
 260 265 270
 Ser Phe Ser Arg Ser Asn Gly Gln Asp Gln Gln Leu Tyr Ile Gly Trp
 275 280 285
 Lys Ser Arg Ser Gly Ser Asp Leu Asp Ala Gly Arg Val Leu Arg Leu
 290 295 300
 Phe Gly Val Asn Ile Ser Pro Glu Ser Ser Arg Asn Asp Val Val Gly
 305 310 315 320
 Asn Lys Arg Val Asn Asp Thr Glu Met Leu Ser Leu Val Cys Ser Lys
 325 330 335
 Lys Gln Arg Ile Phe His Ala Ser
 340

<210> 67

<211> 984

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G659

<400> 67

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 tctctcccca aacaatctgg tatgtcattg cttttgtcat cacaatcaaa gcaaaagcct 180
 cttcaattgt tttttctttt ctttatgatt ctgaatgtat atatatgcaa aaatgaaggg 240
 ctattgaggt gtgggaagag ttgtcgtcta aggtggatta actatcttag gccagatctg 300
 aagcgtggca acttcacttc agaggaggaa gaaacaatca ttaagcttca ccacaactat 360


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gggaacaagt ggtcgaaaat cgcttctcaa cttccaggta gaacagataa cgagatcaag 420
aatgtgtggc aactcatct aaagaaaaga ctggctcaga gctcaggaac tgcagatgaa 480
ccggcctcgc cttgttcgag tgattctgtt tctcgtggga aagatgataa gtcattctcac 540
gtagaagatt ctttgaacag agagactaat cataggaatg agttgtctac atctatgtct 600
tctgggggtt ccaaccaaca agatgatcca aagatagacg aactcagggtt tgagtatata 660
gaagaagctt atagcgagtt taacgacatt attattcaag aggtagacaa acccgatctg 720
ctggagatac ctttgattc agatcctgac atttggagtt tcttagatac ttcaaactca 780
tttcaacaat ccactgcaaa tgagaacagc tcagggtcaa gagcaacaac agaagaagag 840
tctgatgagg atgaggttaa gaaatggttc aagcacctag aaagcgaact cgggttagaa 900
gaagacgata atcaacaaca atacaaagaa gaagaatcat catcatcatc actcttgaag 960
aactacgagc tcatgatata ttga                                     984

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<210> 68

<211> 327

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G659

<400> 68

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Met Gly Lys Gly Arg Ala Pro Cys Cys Asp Lys Thr Lys Val Lys Arg
  1                               10                      15

Gly Pro Trp Ser Pro Glu Glu Asp Ile Lys Leu Ile Ser Phe Ile Gln
  20                      25                      30

Lys Phe Gly His Glu Asn Trp Arg Ser Leu Pro Lys Gln Ser Gly Met
  35                      40                      45

Ser Leu Leu Leu Ser Ser Gln Ser Lys Gln Lys Pro Leu Gln Leu Phe
  50                      55                      60

Phe Leu Phe Phe Met Ile Leu Asn Val Tyr Ile Cys Lys Asn Glu Gly
  65                      70                      75                      80

Leu Leu Arg Cys Gly Lys Ser Cys Arg Leu Arg Trp Ile Asn Tyr Leu
  85                      90                      95

Arg Pro Asp Leu Lys Arg Gly Asn Phe Thr Ser Glu Glu Glu Glu Thr
  100                      105                      110

Ile Ile Lys Leu His His Asn Tyr Gly Asn Lys Trp Ser Lys Ile Ala
  115                      120                      125

Ser Gln Leu Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Val Trp His
  130                      135                      140

Thr His Leu Lys Lys Arg Leu Ala Gln Ser Ser Gly Thr Ala Asp Glu
  145                      150                      155                      160

Pro Ala Ser Pro Cys Ser Ser Asp Ser Val Ser Arg Gly Lys Asp Asp
  165                      170                      175

Lys Ser Ser His Val Glu Asp Ser Leu Asn Arg Glu Thr Asn His Arg
  180                      185                      190

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Asn Glu Leu Ser Thr Ser Met Ser Ser Gly Gly Ser Asn Gln Gln Asp
 195 200 205
 Asp Pro Lys Ile Asp Glu Leu Arg Phe Glu Tyr Ile Glu Glu Ala Tyr
 210 215 220
 Ser Glu Phe Asn Asp Ile Ile Ile Gln Glu Val Asp Lys Pro Asp Leu
 225 230 235 240
 Leu Glu Ile Pro Phe Asp Ser Asp Pro Asp Ile Trp Ser Phe Leu Asp
 245 250 255
 Thr Ser Asn Ser Phe Gln Gln Ser Thr Ala Asn Glu Asn Ser Ser Gly
 260 265 270
 Ser Arg Ala Thr Thr Glu Glu Glu Ser Asp Glu Asp Glu Val Lys Lys
 275 280 285
 Trp Phe Lys His Leu Glu Ser Glu Leu Gly Leu Glu Glu Asp Asp Asn
 290 295 300
 Gln Gln Gln Tyr Lys Glu Glu Glu Ser Ser Ser Ser Ser Leu Leu Lys
 305 310 315 320
 Asn Tyr Glu Leu Met Ile His
 325

<210> 69
 <211> 826
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G620

<400> 69
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 cgtgagcaag accaatacat gccaatcgca aacgtcataa gaatcatgcg taaaacctta 180
 ccgtctcacg ccaaaatctc tgacgacgcc aaagaaacga ttcaagaatg tgtctccgag 240
 tacatcagct tcgtgaccgg tgaagccaac gagcgttgcc aacgtgagca acgtaagacc 300
 ataactgctg aagatatcct ttgggctatg agcaagcttg gggttcgataa ctacgtggac 360
 cccctcacg tgttcattaa ccggtaccgt gagatagaga ccgatcgtgg ttctgcactt 420
 agaggtgagc caccgtcgtt gagacaaacc tatggaggaa atggtattgg gtttcacggc 480
 ccatctcatg gcctacctcc tccgggtcct tatggttatg gtatgttgga ccaatccatg 540
 gttatgggag gtggtcggta ctaccaaacc gggtcgctcg gtcaagatga atccagtgtt 600
 ggtgggtggc cttcgtcttc cattaacgga atgccggctt ttgaccatta tggtcagtat 660
 aagtgaagaa ggagttattc ttcatTTTTA tatctattca aaacatgtgt ttcgatagat 720
 attttatttt tatgtcttat caataacatt tctatataat gttgcttctt taaggaaaag 780
 tgttgtatgt caatacttta tgagaaactg atttatatat gcaaat 826

<210> 70
 <211> 208
 <212> PRT
 <213> Arabidopsis thaliana

<220>

<223> G620

<400> 70

Met Thr Ser Ser Val Ile Val Ala Gly Ala Gly Asp Lys Asn Asn Gly
 1 5 10 15

Ile Val Val Gln Gln Gln Pro Pro Cys Val Ala Arg Glu Gln Asp Gln
 20 25 30

Tyr Met Pro Ile Ala Asn Val Ile Arg Ile Met Arg Lys Thr Leu Pro
 35 40 45

Ser His Ala Lys Ile Ser Asp Asp Ala Lys Glu Thr Ile Gln Glu Cys
 50 55 60

Val Ser Glu Tyr Ile Ser Phe Val Thr Gly Glu Ala Asn Glu Arg Cys
 65 70 75 80

Gln Arg Glu Gln Arg Lys Thr Ile Thr Ala Glu Asp Ile Leu Trp Ala
 85 90 95

Met Ser Lys Leu Gly Phe Asp Asn Tyr Val Asp Pro Leu Thr Val Phe
 100 105 110

Ile Asn Arg Tyr Arg Glu Ile Glu Thr Asp Arg Gly Ser Ala Leu Arg
 115 120 125

Gly Glu Pro Pro Ser Leu Arg Gln Thr Tyr Gly Gly Asn Gly Ile Gly
 130 135 140

Phe His Gly Pro Ser His Gly Leu Pro Pro Gly Pro Tyr Gly Tyr
 145 150 155 160

Gly Met Leu Asp Gln Ser Met Val Met Gly Gly Gly Arg Tyr Tyr Gln
 165 170 175

Asn Gly Ser Ser Gly Gln Asp Glu Ser Ser Val Gly Gly Gly Ser Ser
 180 185 190

Ser Ser Ile Asn Gly Met Pro Ala Phe Asp His Tyr Gly Gln Tyr Lys
 195 200 205

<210> 71

<211> 1394

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G596

<400> 71

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 attagggttt caattgttta ctttttgttt gctttttata tcaagtaatg gatcagggtct 180
 ctgctctctt tcctccacct tttctctcaa gagatctcca tcttcacca caccatcaat 240
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atcacatcac gagaaggcca cgtggcagac cagcgggatc taagaacaaa ccaaaaccgc 480
caatcatcat cactcgagac agcgcaaacg ctctcaaadc tcatgtcatg gaagtagcaa 540
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gcgttttgag cggaaacggc gccgttacca acgttaccat aagacaacca gcttcagtag 660
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gatcattcct tctctctcgg gctccaccag ctgcgtcagg tctaacgatt tacttagccg 780
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gtgttggtga taaa 1394

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<210> 72
<211> 317
<212> PRT
<213> Arabidopsis thaliana

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<220>
<223> G596

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<400> 72
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Leu His Leu His Pro His His Gln Phe Gln His Gln Gln Gln Gln Gln
    20              25              30

Gln Gln Asn His Gly His Asp Ile Asp Gln His Arg Ile Gly Gly Leu
    35              40              45

Lys Arg Asp Arg Asp Ala Asp Ile Asp Pro Asn Glu His Ser Ser Ala
    50              55              60

Gly Lys Asp Gln Ser Thr Pro Gly Ser Gly Gly Glu Ser Gly Gly Gly
    65              70              75              80

Gly Gly Gly Asp Asn His Ile Thr Arg Arg Pro Arg Gly Arg Pro Ala
    85              90              95

Gly Ser Lys Asn Lys Pro Lys Pro Pro Ile Ile Ile Thr Arg Asp Ser
   100              105              110

Ala Asn Ala Leu Lys Ser His Val Met Glu Val Ala Asn Gly Cys Asp
   115              120              125

Val Met Glu Ser Val Thr Val Phe Ala Arg Arg Arg Gln Arg Gly Ile
   130              135              140

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<400>	73						
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agtgatgact	caatgcatcg	tgtcattccc	gtacttgacg	tctttgaggt	cgagcctagt	180	
catcttccaa	atgttgctgg	agtgagatgt	cgaggagacg	ctgagcaatg	gttctttcttc	240	
gtgccacgac	aagaacgcga	agcaagagga	ggcagaccga	gtagaactac	tgggttcagga	300	
tactggaaag	caactggatc	acctgggtcca	gtctttttcca	aagacaacaa	aatgatctga	360	
gcaaaagaaa	ctatggtttt	ctacactgga	aaagcaccga	caggaagaaa	aactaaatgg	420	
aaaatgaatg	agtaccacgc	cgttgacgaa	acagtcaacg	cttcacaaat	ccctaagctg	480	
agacgtgagt	tcagtttatg	tcgagtctac	ataacaacag	gaagctccag	agcttttgat	540	
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cgatactaaa ccagggggga atcaaatga gctgatttcg gaaaaccaag tagagagctt 480
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ttatttcccg gccggtttgc tcaactatta atatgctgct aatttacttg ttttttactt 660
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<211> 171

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G25

<400> 82

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Pro Val Ser Val Ser Glu Glu Arg Asp Gly Lys Arg Glu Arg Lys Asn
      35             40             45

Leu Tyr Arg Gly Ile Arg Gln Arg Pro Trp Gly Lys Trp Ala Ala Glu
      50             55             60

Ile Arg Asp Pro Ser Lys Gly Val Arg Val Trp Leu Gly Thr Phe Lys
      65             70             75             80

Thr Ala Asp Glu Ala Ala Arg Ala Tyr Asp Val Ala Ala Ile Lys Ile
      85             90             95

Arg Gly Arg Lys Ala Lys Leu Asn Phe Pro Asn Thr Gln Val Glu Glu
      100            105            110

Glu Ala Asp Thr Lys Pro Gly Gly Asn Gln Asn Glu Leu Ile Ser Glu
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Asn Gln Val Glu Ser Leu Ser Glu Asp Leu Met Ala Leu Glu Asp Tyr
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Met Arg Phe Tyr Gln Ile Pro Val Ala Asp Asp Gln Ser Ala Thr Asp
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002230: 002230: 002230

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165 170

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<213> Arabidopsis thaliana

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<223> G610

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<213> Arabidopsis thaliana

<223> G610

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35 40 45

Ala Ser Leu Ala Gly Lys Leu Leu Glu Glu Ser Glu Ser Ser Ser Thr
50 55 60

Ser Thr Tyr Ala Ser Glu Ala Asp Asn Leu Asp His Leu Gly Gly Leu
65 70 75 80

Ile Lys Gln Glu Leu Glu Asp Gly Tyr Thr Thr Lys Pro Cys Lys Ser
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Glu Phe Phe Asp Pro Gly Asn Pro Ala Ser Lys Ser Thr Ser Glu Asn
100 105 110

Thr Ser Val Thr Cys Leu Pro Phe Ser Ser Phe Glu Asn Asp Cys Ile
115 120 125

Leu Glu Gln Thr Pro Val Ser Asp Cys Lys Arg Ala Ser Gly Leu Lys
130 135 140

Ser Leu Val Gly Ser Ile Thr Glu Glu Thr Cys Val Val Asn Glu Asp
145 150 155 160

Ala Gly Ser Glu Gln Gly Ala Asn Thr Phe Ser Leu Lys Asp Pro Ser
165 170 175

Gln Leu His Ser Gln Ser Pro Glu Ser Val Leu Leu Asp Gly Asp Val
180 185 190

Lys Leu Ala Pro Cys Thr Asp Gln Val Pro Asn Asp Ser Phe Lys Gly
195 200 205

Tyr Arg Asn His Ser Lys Leu Val Cys Arg Asp Asp Asp Glu Asn Tyr
210 215 220

Cys Lys Tyr Tyr Lys Phe Ser Asp Lys Cys Lys Ser Tyr Arg Pro Leu
225 230 235 240

Ser Arg Val Gly Asn Arg Arg Ile Met Gln Ser Val Arg Ala Ile Ser
245 250 255

Lys Leu Lys Cys Phe Glu Asp Thr Arg Thr Asp Gly Arg Leu Lys Ala
260 265 270

Leu Tyr Arg Lys Arg Lys Leu Cys Tyr Gly Tyr Asn Pro Trp Lys Arg
275 280 285

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Lys	Gly	Glu	Ser	Glu 325	Asn	Gly	Asp	Phe	Ser 330	Ala	Ala	Lys	Ile	Gly 335	Leu
Leu	Ser	Lys	Asp 340	Ser	Arg	Val	Lys	Phe 345	Ser	Ile	Lys	Ser	Leu 350	Arg	Ile
Pro	Glu	Leu 355	Val	Ile	Glu	Val	Pro 360	Glu	Thr	Ala	Thr	Val 365	Gly	Leu	Leu
Lys	Arg 370	Thr	Val	Lys	Glu	Ala 375	Val	Thr	Ala	Leu	Leu 380	Gly	Gly	Gly	Ile
Arg 385	Ile	Gly	Val	Leu	Val 390	Gln	Gly	Lys	Lys	Val 395	Arg	Asp	Asp	Asn	Asn 400
Thr	Leu	Ser	Gln	Thr 405	Gly	Leu	Ser	Cys	Arg 410	Glu	Asn	Leu	Gly	Asn 415	Leu
Gly	Phe	Thr	Leu 420	Glu	Pro	Gly	Leu	Glu	Thr	Leu	Pro	Val	Pro	Leu	Cys
Ser	Glu	Thr	Pro	Val	Leu	Ser	Leu 440	Pro	Thr	Asp	Ser	Thr	Lys	Leu	Ser
Glu 450	Arg	Ser	Ala	Ala	Ser	Pro 455	Ala	Leu	Glu	Thr	Gly 460	Ile	Pro	Leu	Pro
Pro 465	Gln	Asp	Glu	Asp	Tyr 470	Leu	Ile	Asn	Leu	Gly 475	Asn	Ser	Val	Glu	Asn 480
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Leu 530	Ser	Gln	Arg	Arg	Thr	Arg 535	Arg	Leu	Phe	Ser	Val 540	Thr	Glu	Val	Glu
Ala 545	Leu	Val	Ser	Ala	Val 550	Glu	Glu	Val	Gly	Thr	Gly 555	Arg	Trp	Arg	Asp 560
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Gln Gln Arg Arg Gly Glu Pro Val Pro Gln Glu Leu Leu Asp Arg Val
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610 615 620

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<211> 371
<212> PRT
<213> Arabidopsis thaliana

<220>
<223> G229

<400> 86
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Cys	Gly	Lys	Ser	Cys	Arg	Leu	Arg	Trp	Ile	Asn	Tyr	Leu	Arg	Ser	Asp
	50					55					60				
Leu	Lys	Arg	Gly	Asn	Ile	Thr	Pro	Glu	Glu	Glu	Glu	Leu	Val	Val	Lys
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Leu	His	Ser	Thr	Leu	Gly	Asn	Arg	Trp	Ser	Leu	Ile	Ala	Gly	His	Leu
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			100					105					110		
Ser	Arg	Lys	Leu	His	Asn	Phe	Ile	Arg	Lys	Pro	Ser	Ile	Ser	Gln	Asp
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Gln	Ala	Lys	Arg	Arg	Leu	Gly	Arg	Thr	Ser	Arg	Ser	Ala	Met	Lys	Pro
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Lys	Ile	Arg	Arg	Thr	Lys	Thr	Arg	Lys	Thr	Lys	Lys	Thr	Ser	Ala	Pro
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Pro	Glu	Pro	Asn	Ala	Asp	Val	Ala	Gly	Ala	Asp	Lys	Glu	Ala	Leu	Met
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	210					215						220			
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Ser	Asp	Pro	Gly	His	Leu	Tyr	Thr	Asn	Thr	Thr	Cys	Gly	Gly	Gly	Gly
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Glu	Leu	His	Asn	Ile	Arg	Asp	Ser	Glu	Gly	Ala	Arg	Gly	Phe	Ser	Asp
			260					265					270		
Thr	Trp	Asn	Gln	Gly	Asn	Leu	Asp	Cys	Leu	Leu	Gln	Ser	Cys	Pro	Ser
		275					280					285			
Val	Glu	Ser	Phe	Leu	Asn	Tyr	Asp	His	Gln	Val	Asn	Asp	Ala	Ser	Thr
	290					295					300				
Asp	Glu	Phe	Ile	Asp	Trp	Asp	Cys	Val	Trp	Gln	Glu	Gly	Ser	Asp	Asn
305					310					315					320
Asn	Leu	Trp	His	Glu	Lys	Glu	Asn	Pro	Asp	Ser	Met	Val	Ser	Trp	Leu
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<210> 89
<211> 1952
<212> DNA
<213> Arabidopsis thaliana
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<223> "n" bases at various positions throughout the
sequence may be A, T, C, G, other or unknown
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caccgcaggt	tctcaaattcc	tttggcgatg	tctagaattg	acgaagaaga	tgatcagaag 240
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ggcgaagtgg	atttctttctc	cgacaagaaa	cttaggggtt	gtcgtgaaga	cgacgaagga 420
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ctataaatag tncgttnctt antaaaaaaa aa 1952

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<210> 90

<211> 553

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G186

<400> 90

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20 25 30

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35 40 45

Arg Ile Ser Thr Asn Gly Ser Glu Phe Arg Phe Pro Val Ser Leu Ser
50 55 60

Gly Ile Arg Asp Arg Glu Asp Glu Asp Phe Ser Ser Gly Val Ala Gly
65 70 75 80

Asp Asn Asp Arg Glu Val Pro Gly Glu Val Asp Phe Phe Ser Asp Lys
85 90 95

Lys Ser Arg Val Cys Arg Glu Asp Asp Glu Gly Phe Arg Val Lys Lys
100 105 110

Glu	Glu	Gln	Asp	Asp	Arg	Thr	Asp	Val	Asn	Thr	Gly	Leu	Asn	Leu	Arg
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Thr	Thr	Gly	Asn	Thr	Lys	Ser	Asp	Glu	Ser	Met	Ile	Asp	Asp	Gly	Glu
		130				135					140				
Ser	Ser	Glu	Met	Glu	Asp	Lys	Arg	Ala	Lys	Asn	Glu	Leu	Val	Lys	Leu
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Gln	Asp	Glu	Leu	Lys	Lys	Met	Thr	Met	Asp	Asn	Gln	Lys	Leu	Arg	Glu
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			180					185					190		
Val	Ser	Leu	Met	Gln	Gln	Gln	Gln	Gln	Gln	Asn	Asn	Lys	Val	Ile	Glu
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Ala	Ala	Glu	Lys	Pro	Glu	Glu	Thr	Ile	Val	Pro	Arg	Gln	Phe	Ile	Asp
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Leu	Gly	Pro	Thr	Arg	Ala	Val	Gly	Glu	Ala	Glu	Asp	Val	Ser	Asn	Ser
225					230					235					
Ser	Ser	Glu	Asp	Arg	Thr	Arg	Ser	Gly	Gly	Ser	Ser	Ala	Ala	Glu	Arg
				245					250						
Arg	Ser	Asn	Gly	Lys	Arg	Leu	Gly	Arg	Glu	Glu	Ser	Pro	Glu	Thr	Glu
			260					265					270		
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		275					280								
Gln	Thr	Ala	Glu	Ala	Thr	Met	Arg	Lys	Ala	Arg	Val	Ser	Val	Arg	Ala
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Arg	Ser	Glu	Ala	Pro	Met	Ile	Ser	Asp	Gly	Cys	Gln	Trp	Arg	Lys	Tyr
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His	Pro	Leu	Pro	Pro	Ala	Ala	Val	Ala	Met	Ala	Ser	Thr	Thr	Thr	Ala
		370				375									
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Ala Arg Arg Ser Arg Leu Arg Lys Gln Ala Glu Thr Glu Glu Leu Ala
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Arg Lys Val Glu Ala Leu Thr Ala Glu Asn Met Ala Leu Arg Ser Glu
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Thr Leu Leu Asp Lys Leu Lys Cys Ser Glu Pro Glu Lys Arg Val Pro
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<212> DNA

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<223> G255

<400> 93

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 Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Tyr Trp Asn Thr His Ile
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 Lys Arg Lys Leu Leu Ser Lys Gly Ile Asp Pro Ala Thr His Arg Gly
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Gly	Ile	Arg	Arg	Arg	Lys	Trp	Gly	Lys	Trp	Val	Ala	Glu	Ile	Arg	Glu
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Ala	Ala	Ala	Arg	Ala	Tyr	Asp	Val	Ala	Val	Phe	Tyr	Leu	Arg	Gly	Pro
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Ser	Ala	Arg	Leu	Asn	Phe	Pro	Asp	Leu	Leu	Leu	Gln	Glu	Glu	Asp	His
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Lys Ala Ala Glu Val Gly Ala Arg Val Asp Ala Leu Leu Ala Ser Ala
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Gln Gln Leu Ile Arg Glu Cys Pro Ile Leu Cys Asn Ile Glu Pro Arg
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Gln Ile Lys Val Trp Phe Gln Asn Arg Arg Cys Arg Glu Lys Gln Arg
65 70 75 80

Lys Glu Ser Ala Arg Leu Gln Thr Val Asn Arg Lys Leu Ser Ala Met
85 90 95

Asn Lys Leu Leu Met Glu Glu Asn Asp Arg Leu Gln Lys Gln Val Ser
100 105 110

Asn Leu Val Tyr Glu Asn Gly Phe Met Lys His Arg Ile His Thr Ala
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 35 40 45
 Gly Val Val Asp Lys Gln Thr Ser Thr Thr Leu Phe Thr Phe Ser Pro
 50 55 60
 Gly Gly Glu Lys Ser Ser Arg Asp Val Pro Lys Pro His Val Ala Phe
 65 70 75 80

Ala Met Gln Ser Ala Cys Phe Glu Phe Gly Phe Ala Gln Pro Met Met
85 90 95

Tyr Thr Lys His Pro His Val Glu Gln Tyr Tyr Gly Val Val Ser Ala
100 105 110

Tyr Gly Ser Gln Arg Ser Ser Gly Arg Val Met Ile Pro Leu Lys Met
115 120 125

Glu Thr Glu Glu Asp Gly Thr Ile Tyr Val Asn Ser Lys Gln Tyr His
130 135 140

Gly Ile Ile Arg Arg Arg Gln Ser Arg Ala Lys Ala Glu Lys Leu Ser
145 150 155 160

Arg Cys Arg Lys Pro Tyr Met His His Ser Arg His Leu His Ala Met
165 170 175

Arg Arg Pro Arg Gly Ser Gly Gly Arg Phe Leu Asn Thr Lys Thr Ala
180 185 190

Asp Ala Ala Lys Gln Ser Lys Pro Ser Asn Ser Gln Ser Ser Glu Val
195 200 205

Phe His Pro Glu Asn Glu Thr Ile Asn Ser Ser Arg Glu Ala Asn Glu
210 215 220

Ser Asn Leu Ser Asp Ser Ala Val Thr Ser Met Asp Tyr Phe Leu Ser
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<223> G1650

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Leu Phe Ile Gln Glu Asp Glu Met Ala Ser Trp Leu His Gln Pro Asn
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Arg Gln Asp Tyr Leu Tyr Ser Gln Leu Leu Tyr Ser Gly Val Ala Ser
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Thr His Pro Gln Ser Leu Ala Ser Leu Glu Pro Pro Pro Pro Pro Arg
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Ala Gln Tyr Ile Leu Ala Ala Asp Arg Pro Thr Gly His Ile Leu Ala
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Glu Arg Arg Ala Glu Asn Phe Met Asn Ile Ser Arg Gln Arg Gly Asn
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Ile Phe Leu Gly Gly Val Glu Ala Val Pro Ser Asn Ser Thr Leu Leu
      165              170              175

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Ser Ser Ala Thr Glu Ser Ile Pro Ala Thr His Gly Thr Glu Ser Arg
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Trp Gly Leu Cys Lys Ala Glu Thr Glu Pro Val Gln Arg Gln Pro Ala
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Thr Glu Thr Asp Ile Thr Asp Glu Arg Lys Arg Lys Thr Arg Glu Glu
 245 250 255

Thr Asn Val Glu Asn Gln Gly Thr Glu Glu Ala Arg Asp Ser Thr Ser
 260 265 270

Ser Lys Arg Ser Arg Ala Ala Ile Met His Lys Leu Ser Glu Arg Arg
 275 280 285

Arg Arg Gln Lys Ile Asn Glu Met Met Lys Ala Leu Gln Glu Leu Leu
 290 295 300

Pro Arg Cys Thr Lys Thr Asp Arg Ser Ser Met Leu Asp Asp Val Ile
 305 310 315 320

Glu Tyr Val Lys Ser Leu Gln Ser Gln Ile Gln Asp Val Leu Asn Gly
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Thr Cys Tyr Asp Ser Thr Asp Asp Val Cys Gly Glu Tyr Thr Thr Thr
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 35 40 45
 Cys Gly Lys Ser Cys Arg Leu Arg Trp Met Asn Tyr Leu Lys Pro Asp
 50 55 60
 Ile Lys Arg Gly Asn Phe Thr Lys Glu Glu Glu Asp Ala Ile Ile Ser
 65 70 75 80
 Leu His Gln Ile Leu Gly Asn Arg Trp Ser Ala Ile Ala Ala Lys Leu
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 ggtggaccaa ctggtcctaa ggtctcttct tctaccttta attactatat tcataacttt 180
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<223> G171

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<213> *Arabidopsis thaliana*

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